

Stability analyses of sunflower (*Helianthus annuus* L.) hybrids for oleic acid and yield traits under multi location trials in Pakistan

Masood Hussain Shah,¹ Saeed Rauf,¹ Shahid Nazir,² Rodomiro Ortiz,³ Abdul Naveed,⁴ Seerat Fatima¹

¹Department of Plant Breeding and Genetics, College of Agriculture, University of Sargodha, Pakistan; ²Agricultural Biotechnology Research Institute, Ayub Agriculture Research Institute, Faisalabad, Pakistan; ³Department of Plant Breeding, Swedish University of Agricultural Sciences, Alnarp, Sweden; ⁴Office of Research Innovations and Commercialization, University of Agriculture, Faisalabad, Pakistan

Highlights

- An important aim for sunflower breeding is to release hybrids with stable high oleic acid, seed yield and oil content.
- Heritability of oleic acid content was medium across testing sites.
- Oleic acid content and seed yield increased at high temperature and low humidity particularly in the spring season.
- Relation of oleic acid content and combining ability suggests selection effectiveness to breed high oleic acid inbreds.
- H1, H4, H5 and H10 were the most promising hybrids due to their high oleic acid content across sites in the spring.

Correspondence: Rodomiro Ortiz, Department of Plant Breeding, Swedish University of Agricultural Sciences, Box 190, SE 23422 Lomma, Sweden. E-mail: rodomiro.ortiz@slu.se

Key words: combining ability; degree days; kernel to seed percentage; seed yield.

Contributions: MHS, worked as a PhD scholar under the project, carried out agronomic evaluations; SN, conducted DNA profiling of hybrids; AN, co-principal investigator of project. SR, principal investigator of the project who conceived the research; RO, made major improvement in the manuscript and technical aspects of research.

Conflict of interest: the authors declare no potential conflict of interest.

Availability of data and materials: data and materials are available by the authors.

Acknowledgements: this study was partly funded by the ALP-PARC project entitled: development of sunflower hybrids modified for high oleic acid in edible oil (C-163).

See online Appendix for additional materials.

Received for publication: 15 March 2022. Accepted for publication: 9 November 2022.

©Copyright: the Author(s), 2023 Licensee PAGEPress, Italy Italian Journal of Agronomy 2023; 18:2079 doi:10.4081/ija.2023.2079

This article is distributed under the terms of the Creative Commons Attribution Noncommercial License (by-nc 4.0) which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

Publisher's note: all claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article or claim that may be made by its manufacturer is not guaranteed or endorsed by the publisher.

Abstract

The development of a hybrid with high oleic acid is an important breeding goal for sunflower. High oleic acid sunflower has better cooking quality due to low oxidation and rancidity. Hence, inbred lines differing for oleic acid content were selected, alongside the development of hybrids where one or both parents exhibited high oleic acid content in edible oil, and then evaluated at various sites (i.e. with comparatively low temperature during sunflower reproductive phase at Sargodha and Faisalabad; while high temperature and low humidity at Bhawalpur and Multan) in Pakistan during spring season. Moreover, autumn season was relatively cool and high humid for sites (Faisalabad and Sargodha, Pakistan). DNA profiling of hybrids differing for oleic acid content using N1-3F/N2-1R confirmed the presence of a high oleic acid allele in the hybrids. Oleic acid content and seed yield components were increased at high temperature and low humidity to a greater extent in spring than in autumn season. Among the hybrids, one (H5) had stable high oleic acid content during the spring season with higher seed yield and kernel to seed percentage than the check cultivars (Hysun-33 and FH-331). Analysis of the combining ability of two locations revealed a relationship between mean oleic acid contents and combining ability, thereby suggesting the effectiveness of selection in developing high oleic acid inbred lines. Newly developed inbred C.112.P was a positive combiner for oleic acid at all sites except Sargodha, while restorer populations such as RH.344, RH.345 and RH.347 were positive male combiners.

Introduction

Sunflower seed is a rich source of edible oil (30-50%), protein (20-30%), tocopehrols (vitamin E), and fatty acids. Its oil contains up to 90% polyunsaturated fatty acid (60% linoleic acid) (Rauf, 2019). Improvement and modification of sunflower oil and fatty acid is an important breeding goal (Alberio *et al.*, 2017). Regular sunflower oil (with high linoleic acid) is more suitable as

a salad dresser. This is partly due to presence of poly-unsaturated fatty acid in sunflower oil. Sunflower oil rich in linoleic acid is ideal for consumption but such oil shows poor oxidative stability as a cooking or deep-frying oil (Chernova et al., 2021). Oil rich in polyunsaturated fatty acids becomes rancid quickly due to its susceptibility to oxidation (Romano et al., 2021). On the other hand, oils enriched in saturated fatty acids or mono-unsaturated fatty acid (oleic acid) show more stability during cooking and have greater shelf life than polyunsaturated fatty acids (Romano et al., 2021). However, saturated fatty acid increases blood cholesterol level and other health hazards (Rauf et al., 2020; Astrup et al., 2021). Fatty acid profiles showed that mid- to high oleic hybrids had oleic acid ranging from 44 to 85%, 9 to 46% linoleic acid, 4 to 6% palmitic acid, and 2 to 6% stearic acid (de Carvalho et al., 2019). Sunflower accessions have been characterized into three types with respect to oleic acid content such as standard sunflower types (<50%), mid oleic acid (50-70%) and high oleic acid content (>80%) (Rauf, 2019; Manalili et al., 2021). High oleic acid content was controlled by a single dominant gene mutation. This dominant mutation was induced in breeding program through use of a chemical mutagen ethyl methane sulfonate (Dimitrijević et al., 2017; Rauf et al., 2017; Rauf, 2019).

The Pervenent genotype (obtained through a mutation breeding program) was selected in breeding programs as a sunflower cultivar characterized by high oleic acid. Oleic acid content in Pervenent was highly unstable and ranged between 15-51% and 87-91% across a range of environments, depending upon temperature during the reproductive phase (Alberio *et al.*, 2017). NuSun has been released for general cultivation in the USA as mid-oleic acid commercial cultivar of sunflower (Gupta, 2014).

Temperature, particularly minimum night temperature, and intercepted solar radiation have positive effects on oleic acid content in sunflower (Echarte *et al.*, 2010). Sowing also affects the oleic acid content of sunflower due to the influence of temperature, humidity and rainfall (Akkaya *et al.*, 2019). Early sowing decreased oleic acid and palmitic acids and increased linoleic and stearic acids under irrigation, and irrigation decreased oleic acid contents in a sunflower study by Flagella *et al.*, (2002). In Pakistan, regular sunflower hybrid crops can be grown in different seasons (spring and autumn). Spring season crops are generally sown in February while autumn season cultivation was recommended in August in Pakistan (Qadir *et al.*, 2006). Both sowing periods differ in intercepted radiation, humidity and rain fall and thus provide different environmental condition for growth and



reproduction associated with oleic acid synthesis (Qadir et al., 2006). Stability analyses have been carried out to select hybrids with high oleic acid content across environments by Van Der Merwe et al. (2013), who evaluated 16 sunflower hybrids and found that one displayed high oleic levels (80%) across all environments. Unstable hybrids may be sensitive to temperature, rainfall and humidity, and will not benefit farmers in some years. In addition to stability across environments, the stability of performance of a parent with multiple other lines to develop good hybrids is essential. Combining ability analyses of 15 hybrids generated from breeding lines differing for oleic acid content revealed dominance or partial dominance in the F₁ for some of the parents (Joksimović et al., 2006). Thus, inbred lines differed in their ability to produce high oleic acid hybrids depending on the other parent; however, some produced high oleic acid hybrids with multiple inbred lines, and were positive general combiners (Joksimović et al., 2006). Oleic acid traits measured in these crosses showed a preponderance of additive gene action, while dominance and epistatic effects were non-significant (Joksimović et al., 2006).

On the basis of this background, the aim of our research was to select inbred lines with high oleic acid for further crossing to develop hybrids, and to evaluate these hybrids in various locations during spring [4 locations: Sargodha, Faisalabad (with comparatively lower temperature than Multan and Bahawalpur across 2 years) and autumn seasons (2 locations with low reproductive phase temperature and higher humidity)] in Pakistan.

Materials and Methods

Development of breeding lines

Pervenent high oleic acid sunflower (89% oleic acid) was introduced from the United States Department of Agriculture (USDA), and superior plants with high fertility were selected and backcrossed to cytoplasmic male sterile lines (CMS-89) to develop two inbred lines C.112.P and C.116.P. High oleic acid restorer populations (RH.345, RH.347 and RH.345) were introduced from the USDA and selected (during year 2018-20) and maintained for purity for several generations. These restorers originally had more than 80% oleic acid content. Low oleic acid inbred lines C.250, C.249, RSIN.82 and R.365 were also included in the crossing. Seeds of the inbred lines were multiplied and produced hybrids as indicated in Table 1. Those hybrids having sufficient seeds were

Tabl	e 1	. (Code	and	salient	feature	of	hy	brid	ls d	leve	loped	and	eva	luated	in	the	stud	ly.
------	-----	-----	------	-----	---------	---------	----	----	------	------	------	-------	-----	-----	--------	----	-----	------	-----

Coue	Tarcinage	Sanchi Icatule
FH.331	Cultivar check	Local hybrid, dwarf and early maturing hybrid, low oleic acid hybrid
Hysun-33	Commercial check	High yielding, tall and late maturing hybrid, low oleic acid hybrid
H1	$C.112.P \times RH.344$	Both high oleic acid parents
H2	$C.112.P \times RSIN.82$	High \times low oleic acid parent
H3	$C.112.P \times RH.365$	High \times low oleic acid parent
H4	$C.250 \times RH.345$	Low \times high oleic acid parent
H5	$C.112.P \times RH.347$	Both high oleic acid parents
H6	$C116.P \times RH.365$	High \times low oleic acid parent
H7	$C.249 \times RH.345$	Low \times high oleic acid parent
H8	C.116.P × RH.344	Both high oleic acid parents
H9	C.250 × RH344	Low \times high oleic acid parent
H10	$C.249 \times RH.447$	Low \times high oleic acid parent

Coliont footuur



evaluated in multisite trials (4 locations: Sargodha, Faisalabad, Bahawalpur and Multan) over spring and autumn seasons. Hybrid seed was produced by growing cytoplasmic male sterile lines (CMS) and restorer (R) lines (2:1). Synchronized plants of CMS and R combinations were bagged together before opening of ray florets to avoid insect pollinators and CMS lines were manually pollinated from R lines to improve seed set. CMS lines were also maintained from their B lines while restorer and maintainer lines were maintained through sib mating by bagging two plants of same inbred lines. Mature hybrid seed was manually threshed, dried and put in paper bags and stored at room temperature.

Evaluation of hybrids

The F₁ hybrids were planted in two locations on 15th August 2020 for evaluation during the autumn season; and at four locations on 16th February 2020 for evaluation during the spring. Details of all locations and meteorological features are given in Table 2 and Supplementary Figure S1. Daily mean minimum and maximum temperature is shown in Figures 1 and 2. Mean monthly temperature (°C) fluctuation during growth cycles of sunflower crop is shown in Tables 3 and 4. Temperature during the spring season was low during the vegetative phase and high during the reproductive phase (Figure 2; Table 3). In contrast, the autumn season had relatively higher temperatures during vegetative phase (Figure 1; Table 4). Among the experimental sites, Multan and

Bahawalpur had very high temperature stress during the reproductive and grain filling stages (April-May) when compared with Sargodha and Faisalabad (Table 3). Sargodha had temperature stress free growth season during spring season (Table 3).

The soil of the Faisalabad site was a sandy clay loam type with a pH of 7.4 \pm 0.27, EC equal to 2.19 \pm 0.1 deci Siemen meter⁻¹ (dS m⁻¹) and a water holding capacity 18.5% by weight determined through the gravimetric method (Reynolds, 1970). Soil at the Faisalabad site contained Potassium (K) levels of 161.3±4.00 mg kg⁻¹, and phosphorous (P⁻) of 8.31±0.58 mg kg⁻¹. The Sargodha site has a sandy loam type of soil with a pH of 7.6±0.11, EC equal to 1.67 \pm 0.08 dS m⁻¹, organic matter 0.82 \pm 0.11, and 17.5% water holding capacity by weight, while K^+ contents were 172.3 \pm 3.17 mg kg⁻¹, and P⁻ 8.39 ± 0.42 mg kg⁻¹. Multan has a loam soil type with a pH of 8.86 ± 0.19 , EC equal to 1.85 ± 1.08 dS m⁻¹, organic matter being 0.42% ± 0.05 , K⁺ contents were 177.5 ± 5.00 mg kg⁻¹, and P- was 7.47±0.72 mg kg⁻¹, and 18.3% water holding capacity by weight. Bahawalpur has a sandy soil with a pH of 7.63±0.09, EC equal to 2.15 \pm 0.22 dS m⁻¹, organic matter 0.64% \pm 0.04, K⁺ of 124.5±7.13 mg kg⁻¹, P⁻ 11.37±1.01 mg kg⁻¹, and 15.2% water holding capacity. Each hybrid was sown in three rows of 6 m following a randomized complete block design with three replications. Each row was about 40 m long, plants to plant distance was maintained at 22 cm while the distance between rows was 75 cm. Soils were fertilized with inorganic fertilizer, 60 Kg ha-1 phospho-

Table 2. Characteristics of the experimental sites used for field trials across seasons in Pakistan.

Trait		Spri	ng		Autu	mn
	Sargodha	Faisalabad	Bahawalpur	Multan	Sargodha	Faisalabad
Location	32.07°N, 72.69°E	31.45°N, 73.14°E	29.35°N, 71.69°E	30.16°N, 71.52°E	32.07°N, 72.69°E	31.45°N, 73.13°E
Elevation (m)	190	186	214	122	190	186
Vegetative days	721	880.5	1406.5	1240	861.5	931.75
Reproductive days	835	969.5	1336.5	1570	830	678
Rainfall (mm)	181.5	170.0	71.6	120.6	43.1	56
Preceding crop	Brassica juncea	Brassica juncea	Brassica juncea	Brassica juncea	Triticum aestivum	Triticum aestivum
Supplemental irrigation	1.0 (76 mm)	1.0 (76 mm)	4.0 (304 mm)	4.0 (304 mm)	1.0 (76mm)	1.0 (76mm)
Relative humidity	52	62	29	39	68	73

T11 2 11	.11	. (0		- 6	1 .	. 1		•	• .	1 •	•	
Table 5 Mon	nthiv tem	neratiire (*		- t	luctuation amo	na th		neriment	SITES A	11111100	chrine	r ceacon
1abic J. 1101	iuniy iuni	perature (\mathbf{v}		iuciation amo	1 <u>2</u> 11	н сл	perment	SILCS V	aurme	SPILLE	scasom.
		· · · · · · · · · · · · · · · · · · ·										,

Month M		tan	Bahav	valpur	Faisa	alabad	Sargo	Sargodha		
	Max	Min	Max	Min	Max	Min	Max	Min		
Feb (sowing)	26.20 ± 4.60	15.60 ± 2.87	29.80 ± 3.71	15.93 ± 2.05	24.56 ± 2.34	12.56±2.80	22.81±3.23	10.06 ± 1.53		
March (vegetative)	33.63 ± 4.45	22.10 ± 3.80	32.47 ± 4.46	18.70 ± 3.67	25.22 ± 4.25	14.41 ± 1.19	17.55 ± 4.05	12.44 ± 2.09		
April (anthesis)	41.10 ± 2.90	28.76 ± 2.71	40.34 ± 2.63	25.85 ± 2.49	33.17 ± 3.13	20.57 ± 2.40	29.67 ± 3.50	17.93 ± 2.33		
May (grain filling)	44.96 ± 3.69	34.25 ± 2.41	43.12 ± 3.30	28.65 ± 3.30	38.22 ± 3.87	24.39 ± 2.46	28.47 ± 3.27	21.17 ± 2.72		

Table 4. Monthly temperature (°C) fluctuation among the experiment sites during autumn season.

Month	Sargo	dha	Faisalabad					
	Max	Min	Max	Min				
August (sowing)	34.81±1.47	26.06 ± 1.76	39.41±1.25	28.88±1.41				
September (vegetative)	29.85 ± 3.20	22.78 ± 2.89	34.22 ± 2.45	23.23 ± 3.40				
October (anthesis)	29.57 ± 1.50	16.57 ± 2.33	33.22±3.14	15.78 ± 2.51				
November (grain filling)	14.70 ± 2.53	12.04 ± 1.77	22.83 ± 2.48	10.17 ± 2.38				















rus [6 bags (each 50 Kg) of diammonium phosphate, Engro, Pakistan], and 40 kg ha⁻¹ of Nitrogen [2 bag (each 50 kg) of urea, Engro, Pakistan] during field preparation. The pre-emergence herbicide S-metolachlor (dual gold, Syngenta, Pakistan) was sprayed at all sites to control the growth of weeds. Pest scouting was done throughout, and recommended pesticide was sprayed (20 mL dissolved in 20 L water) when armyworm (Coragen, FMC, Pakistan) populations reached above threshold levels during the spring season. No specific infestation was seen at either site during the autumn season.

Hybrid seed production for on farm yield trials

Inbred lines were planted during the 2021 autumn season (15th August 2021) for mass production of hybrid seed. Inbred lines (C-112.P, C-116P, C.250, RSIN.82, RH.344, RH.347, RH.345) were sown as (2:1) of female and male parents to yield several cross combinations. Cross combinations were obtained by bagging each of the floral head as bud was of sufficient size before opening. All unbagged buds were removed before their opening. Each male line was sown in the middle of two CMS lines. Pollen was transferred manually from the bagged male floral heads by shaking male head over CMS heads. The procedure was continued until all the stigma withered and seed started to develop within floral head. Mature floral heads were harvested and dried under the shade. Seeds were manually threshed, cleaned and dried, and put in a Kraft paper bag and stored at room temperature.

On farm yield trials

Spring season sunflower yield trials were performed at four locations (Shahkot, Sargodha, Multan and Bahawalpur) with seeds sown on 12th February 2021. About 100 g seed of each hybrid (H1, H2, H4, H5, H8, H9 and Hysun.33) were given to farmers for plantation in the field on ridges with plant-to-plant distance of 22 cm. Each hybrid was sown in 12 rows of 67 m, which were 75 cm apart.

Measurement of seed yield and components

Five heads from consecutive plants within each of 3 rows (I m⁻²) were harvested and threshed manually. Seed from all hybrids from each replication within each location were dried to a constant moisture (8%) under natural sunlight. Seed yield (g m⁻²) was determined on a digital balance. Seed lots of 100 g were obtained after de-hulling using a rotary machine. The mass of de-hulled seed was determined to calculate kernel to achene ratio as follows:

Kernel to achene ratio = mass of kernel after dehulling (g) \times 100 mass of 100 g of achene

Kernel oil content after de-hulling was determined on a soxhlet apparatus (64826, Sigma, USA). About 10 g of kernels were crushed and put in thimble to extract oil through petroleum ether. Kernel oil contents were determined by following equation:

Kernel oil contents (%) = [KM before extraction (g) – KM after extraction (g)] × 100 KM before extraction

where KM is kernel mass (g)

Fatty acid profile

A manual hand oil extractor (hand extractor, locally assembled, Pakistan) was used to obtain a small quantity of oil (1.5 g) without applying heat. The oil was put in Eppendorf tubes to pro-

file the fatty acids. Small amounts of seed (10 g) were put in the extractor and pressed to obtain about 1.5 g of oil. A 50 μ L sample of this oil was methylated using 4 mL KOH for one hour at room temperature. Methylated fatty acids were extracted with hexane. Fatty acid profiles of all edible oils were analyzed using gas chromatography (M-3900, Varian, USA). Analysis was done using the fused capillary column, flame ionizing detector and nitrogen gas carrier at 3.5 mL min⁻¹. Injector and detector temperature were set at 260°C, while column oven temperature was set at 222°C. Methylated esterified fatty acids were injected manually while fatty acids were identified through peak retention time when compared with a known standard (Sigma Aldrich, USA, purity \geq 98%).

DNA profiling

DNA of sunflower hybrid seed obtained after crossing among various parental lines were amplified with primer N1-3F/N2-1R. This primer has been extensively utilized for identification of high oleic acid sunflower germplasm (Bervillé et al., 2009; Dimitrijević et al., 2017; Bilgen et al., 2018). DNA profiling was carried out by the Agricultural Biotechnology Research Institute, Pakistan. Sunflower seed samples of various hybrids were first frozen in liquid nitrogen and ground into a fine powder. About 200 mg of the samples were used to isolate the total genomic DNA using cetyltrimethylammonium bromide methodologies as described by Rogers and Bendich (1985) with some modifications. Briefly, about 100 ng of isolated DNA was used in polymerase chain reaction (PCR) for validation of the presence of the allele causing high oleic acid using specific primers i.e. N1-3F/N2-1R given by Bilgen et al., 2018. The PCR profile was comprised of 35 cycles of 95°C for 30 seconds, 56°C for 45 seconds and 72°C for 1 minute with a final step of 72°C for 7 minutes. The amplified PCR product was separated on 1.5% agarose gel and visualized under UV gel documentation system.

Statistical and biometrical analyses

All data were analyzed using the computer-based R software (Ferreira et al., 2014) considering a randomized complete block design with 3 replications at 4 locations during the spring season, while 3 replications in each of the 2 locations were used for the autumn season. R functions GGEbiplotGUI were used for GGE biplot analysis, which was originally outlined by Yan and Kang (2002). Stability parameters such as ecovalence (W_i^2 ; Wricke, 1962), regression coefficient (b_i; Finlay and Wilkinson, 1963), deviation from regression (s²d_i; Eberhart and Russel, 1966), stability variance (σ_{i}^{2} ; Shukla, 1972), coefficient of variation (CV, Francis and Kannenberg, 1978); mean and genotype × environment variance component, ($\theta_{(i)}$ and θ_i : Plaisted and Peterson, 1959), and ranking (KR; Kang, 1988) were estimated using STA-BILITYSOFT (Pour-Aboughadareh et al., 2019). The estimated eco-valence (wi²) is the contribution of each genotype to its interaction with the environment. A low value indicates trait stability. The regression coefficient (b_i) also allows identification of stable genotypes. Combining ability effects were estimated on an Excel worksheet following Kempthorne (1957). Mean values of traits were compared using least significant differences at a probability threshold below or equal to 0.05.

Results

PCR identified the high oleic allele of 870 bp in medium and high oleic acid sunflower hybrids (2, 3, 4 and 6; lane



 $2=C.112.P\times R.SIN.82$ (H2); $3=C.112.P\times RH.344$; $4=C.249\times RH.344$; $6=C.250\times RH.344$; while the high oleic acid band was absent in other hybrids (lane 1=Hysun.33, 5=C.224×R.SIN.82; 7=FH.33) (Figure 3).

There were significant ($P \le 0.05$) differences among the hybrids, locations and the interaction of hybrids × locations according to the analysis of variance (Table 5). The significant ($P \le 0.05$) hybrids × location interaction indicated that hybrids changed their relative performance across locations. Hence, means were compared with reference to their specific location. Oleic acid had a medium heritability estimate, while kernel oil contents percentage (KOC%) and seed yield (SY) had low heritability estimates across four locations during the spring season (Table 5).

Among inbred lines B.250, B.249 and R.SIN.82 had the highest de-hulling percentage (KTS%). while RH.344 had the highest KOC%. Oleic acid content of parental lines during the spring season are given in Table 6. Parental lines B.116.P and B.112.P, which were selected from the Pervenent population, had the highest oleic acid content as female lines. Introduced restorer populations produced 70 to 75% oleic acid averaged across the four locations (Table 6).

The hybrids had the highest SY (g m⁻²) at Multan. Among the hybrids, H5 had the highest seed yield at Multan and Faisalabad. Lowest SY values were noted at Sargodha during the spring season (Figure 4A), where hybrid H1 had the highest SY. Hybrid H3 had the highest SY at Bahawalpur (Figure 4A). With respect to location responses for KOC% and kernel to seed percentage (KTS%), mean values of these traits were highest at Sargodha, where on average the lowest SY was observed during the spring season. Hybrids FH.331, H3, H5 and H10 had the highest KTS% at Sargodha, while H4 had the highest KTS% at Multan. H1 had the highest KTS% at Bahawalpur, and FH.331 at Faisalabad during spring season (Figure 4B). Hybrid FH.331 (check cultivar) and hybrids H1, H2, H3 and H7 had the highest KOC% at Sargodha (Figure 4A).



Figure 3. Amplification 870 bp fragment in lanes 2, 3, 4 and 6 that are specific for high oleic (HO), thus confirming the successful cross combinations for HO in sunflower. Lane 1=Hysun.33, 2=C.112.P×R.SIN.82 (H2); 3=C.112.P×RH.344; 4=C.249×RH.344; 5=C.224×R.SIN.82; 6=C.250×RH. 344; 7=FH.331.

Sources of verticition	Deguase of fundam		Moon of	110 400	
Sources of variation	Degrees of freedom	Oil contents (g)	Oleic acid (%)	Kernel to seed (%)	Seed yield (g m ⁻²)
Blocks	2	1.30 ^{NS}	70.05NS	3.60 ^{NS}	377.55NS
Hybrids (H)	11	35.38**	1256.37**	51.64**	16088.46**
Locations (L)	3	183.26**	1607.93**	456.04**	438595.16**
H×L	33	22.97**	203.18**	52.01**	5863.14**
Residual	94	3.16	35.20	9.29	1254.55
Total	143	13.96	201.38	31.70	12621.85
σ²genotype		1.03	87.77		852.11
σ^2 phenotype		10.80	176.03		3642.86
Heritability (h ²)		0.10	0.49		0.23

Table	5.	Analyses	of vari	ance fo	or seed	vield	l and	qualit	y traits	during	spring	season in	1 sunflowe	r and	l estimated	variances	(σ^2)).
						1											· · ·	e

**, $^{\text{NS}}$ indicate significant at P≤0.01 and non-significant at P>0.05, respectively.



H1 had the highest KOC% at Faisalabad, Bahawalpur and Multan (Figure 5A). There were significant differences ($P \le 0.05$) among the hybrids for oleic acid (%) at all locations and seasons. Both commercial hybrids had lower oleic acid percentage at all locations (Figure 5B). The highest oleic acid was obtained at Multan and Bahawalpur while hybrids showed the lowest values of oleic

acid% at Sargodha and Faisalabad during the spring season. H1 and H5 had the highest oleic acid percentage ($\geq 80\%$) at Bahawalpur, while hybrid H9, H2 and H5 ($\geq 80\%$) had the highest oleic acid ($\geq 70\%$) at Multan. H1 had the highest oleic acid percentage at ($\geq 70\%$) at Faisalabad while H7 and H8 showed the highest oleic acid percentage ($\geq 60\%$) at Sargodha (Figure 5B). Multan and



Figure 4. Characteristics of newly developed hybrids along with cultivar checks in the spring season. A) Seed yield (g m-2); B) de-hulling percentage (kernel to seed percentage).

Table 6. Average seed of	quality traits of	f various ma	intainer and	l restorer	lines used	l for the	development	of hybrids to	be grov	vn in the
spring season.							-	-	U	

Parents	Kernel to seed (%)	Kernel oil content (%)	Oleic acid (%)
B.208	56.00 ± 3.18	43.10 ± 1.74	35.55 ± 1.47
B.116.P	36.40 ± 2.27	40.12±2.21	71.24±1.52
R.SIN.82	62.00 ± 2.38	45.34 ± 1.92	35.55 ± 1.67
RH.344	56.40 ± 4.15	52.22 ± 1.34	70.00 ± 2.12
B.250	68.40 ± 1.67	44.11±2.12	52.31 ± 2.33
RH.345	51.00 ± 2.93	42.15 ± 1.69	69.27 ± 1.22
RH.365	55.60 ± 3.71	44.10 ± 3.54	40.93 ± 1.39
B.112.P	48.00 ± 3.57	43.19±3.19	78.38±4.12
B.249	63.20 ± 4.19	45.33 ± 2.54	44.09 ± 2.19
RH.347	61.20 ± 3.38	50.13 ± 1.69	75.23 ± 2.16
RH.447	53.24 ± 4.19	47.21 ± 1.86	74.29 ± 3.19



Bahawalpur had favorable environmental conditions for SY and oleic% during the spring season. Sargodha had the highest oleic acid percentage in comparison to Faisalabad during the autumn season. Hybrid H10 had the highest oleic acid percentage ($\geq 60\%$) at Sargodha and Faisalabad followed by H3 at both locations (Figure 6A). FH.331 and H10 showed similar SY values across both locations during the autumn season. Hybrid H7 had the highest SY during the autumn season followed by H2 and H3. H10 had the highest SY at Faisalabad during the autumn season (Figure 6B). Biplot analysis of oleic acid% showed that H4, H5 and H10 had relatively stable oil content across locations during the spring season (Figure 7). FH.331 had stable but low oleic acid %. H1 had the highest oleic acid at Multan and Bahawalpur during the spring season. Although Hysun.33 was characterized as being a low oleic acid hybrid, but it produced comparatively higher oleic acid at Multan than in other locations (Figure 7).

Stability parameters estimated for oleic acid percentage and seed yield are given in Tables 7 and 8, respectively. Hybrid H5 had

the highest oleic acid contents across locations (Table 7), followed by H7 and H8 during spring season. As per wi², H10 (mid-oleic acid%) had the lowest wi² value followed by H1 (high oleic acid hybrid). H7 and H10 had the smallest Shukla's σ_i^2 stability values, thus being stable. Genotypes with b_i about 1 and non-significant S_{di}^2 were regarded as stable. Hybrids H10, H4 and H5 had regression coefficient about 1 with lowest non-significant S_{di}^2 , and high SY. The hybrids H5, H4, H7 and H10 were the most stable and with the highest oleic acid content.

Hybrid H5, H1 and H6 had the highest SY (g m⁻²) across locations (Table 8). H8 had the lowest eco-valence and Shukla's σ_{i}^2 . H7, H1 and H10 had b_i about 1. H8 had a non-significant S_{di}^2 that confirms its stability for SY. Hybrid H8 also had the lowest Kang's ranking (KR), which indicates that this hybrid as having the highest stability along with high SY when grown in the spring season.

Combining ability effects showed significant ($P \le 0.05$) relationships with mean oleic acid contents of inbred lines at Faisalabad and Sargodha. The variability among inbred lines was



Figure 5. Characteristics of newly developed hybrids along with cultivar checks in the spring season. Bars showing similar alphabets are statistically insignificant (P≥0.05) estimated by least significant difference test. A) Kernel oil content (%); B) oleic acid percentage.



masked by the environment at both Sargodha and Multan, where relationship between combining ability effects and mean oleic acid contents were not high (Figure 8). Among inbred lines, RH>347, RH.344 and B.112.P had higher combining ability values for oleic acid content across locations except Sargodha, where RH.347 and RH.345 were positive combiners. B.112.P had high mean oleic acid content but with negative combining ability effects (Figure 8).

Analysis of variance showed significant variation due to hybrids and hybrids \times locations. Hybrid H2 (oleic acid contents 75.01%) had the highest seed yield averaged over all locations followed by H1 (oleic acid content 81.38%) and standard check (oleic acid contents 35.06%) (Figure 9). Hybrids had the highest seed yield at the Multan where Hysun.33 (oleic acid contents 35.06%) and H2 (oleic acid contents 80.5%) showed the highest seed yield potential (Figure 9). H8 (oleic acid content 76.03%) and H1 (oleic acid content 82.6%) had the highest yield at Bahawalpur (Figure 9).

Discussion

Our research was initiated to develop high oleic acid hybrids with better SY. Breeding lines with high oleic acid and high fertility were derived from Pervenent. These newly developed breeding lines were further used as female lines in the breeding program. They were mated with low oleic or high oleic acid restorer populations. DNA profiling of the hybrids provided a low cost and reliable method of identification of high oleic acid hybrids. Field screening of the high oleic acid sunflower is complicated due to instability of the gene at high temperature as identified in our research. The high oleic acid allele was detected in sunflower hybrids that were successful in breeding programs to develop high oleic acid breeding lines. The developed hybrids were evaluated at four locations, which differed for the total degree days received during their reproductive phase.

Generally, those hybrids whose parents were selected on the

Table 7. Oleic acid contents % and stability parameters ^Z estimated from four locations in Pakistan during spring season.

Hybrid	Y	\mathbf{W}_{i}^{2}	σ^{2}_{i}	s²d _i	b _i	CVi	θ _(i)	$\boldsymbol{\theta}_{i}$	KR
Hysun-33	53.96	446.94	172.00	2.18	-0.79	10.70	58.25	118.51	21
FH.331	42.45	55.25	15.33	5.20	0.63	12.81	72.49	47.30	17
H1	71.01	484.03	186.84	54.77	1.87	23.71	56.90	125.25	16
H2	58.55	607.65	236.29	19.78	2.87	34.77	52.40	147.73	22
H3	67.32	54.86	15.17	2.25	1.54	15.67	72.50	47.22	11
H4	71.76	16.15	-0.31	2.30	1.02	10.03	73.91	40.19	6
H5	79.38	41.59	9.86	5.68	1.12	10.46	72.99	44.81	4
H6	59.06	186.90	67.99	23.02	0.56	13.94	67.70	71.23	18
H7	73.33	74.16	22.89	1.34	0.30	3.68	71.80	50.73	8
H8	72.32	125.15	43.29	11.86	0.44	8.33	69.95	60.00	10
H9	70.14	130.99	45.62	15.37	1.42	15.98	69.74	61.07	14
H10	67.10	11.29	-2.26	1.59	1.03	10.67	74.09	39.30	9

Y, mean seed yield (g m⁻²) across all locations; Wi2, Wricke's (1962) ecovalence; σ²i, Shukla's (1972) stability variance; s²d_i, Finlay and Wilkinson's (1963) deviation from regression; b_i, Eberhart and Russell's (1966) regression coefficient; CV, Francis and Kannenberg's (1978) coefficient of variation; θ_(i) and θ_i, Plaisted and Peterson's (1959) mean and genotype × environment variance component; KR, Kang's (1988) ranking.

TT11 0 C 1	. 11/	\$7	1) 1	1 111		7 . 1	C (r 1		•	D 1 .	1 .	•	
Table 8. Seed y	vield (Y. om	1 ⁻¹) and	stability	parameters	² estimated	trom t	tour l	ocations	111	Pakistan	during	spring	season.
raore or over		-, ,	-)	o etto integ	Paratetter						-		° P	000000

Hybrid	Y	\mathbf{W}_{i}^{2}	σ^{2}_{i}	$s^2 d_i$	bi	CVi	$\theta_{(i)}$	θί	KR
HFH.331	280.17	5587.83	2039.70	744.97	0.90	38.42	1946.58	2090.85	18.00
Hysun-33	321.04	6426.11	2375.01	806.63	1.15	41.66	1916.09	2243.27	16.00
H1	357.63	4941.27	1781.07	703.68	1.02	33.48	1970.09	1973.30	9.00
H2	324.92	5792.15	2121.43	386.23	0.71	25.81	1939.15	2128.00	14.00
H3	297.33	20103.92	7846.14	2104.95	0.62	32.86	1418.72	4730.14	21.00
H4	305.71	6440.79	2380.88	461.98	1.30	48.02	1915.56	2245.94	19.00
H5	393.13	4323.06	1533.79	454.16	1.18	34.07	1992.57	1860.90	7.00
H6	353.92	2489.33	800.30	91.97	1.22	38.42	2059.25	1527.49	6.00
H7	273.17	1608.61	448.01	222.89	1.04	42.70	2091.28	1367.36	14.00
H8	337.29	774.11	114.21	18.15	1.13	37.13	2121.62	1215.63	5.00
H9	318.42	2662.75	869.67	67.03	0.76	26.47	2052.94	1559.02	11.00
H10	275.17	3343.18	1141.84	476.58	0.99	41.37	2028.20	1682.74	16.00

Y, mean seed yield (g m⁻¹) across all locations; Wi2, Wricke's (1962) ecovalence; σ²i, Shukla's (1972) stability variance; s²d_i, Finlay and Wilkinson's (1963) deviation from regression; b_i, Eberhart and Russell's (1966) regression coefficient, θ_(i) and θ(i); CV, Francis and Kannenberg's (1978) coefficient of variation; θ_(i) and θ_i, Plaisted and Peterson's (1959) mean and genotype × environment variance component; KR, Kang's (1988) ranking.



basis of high oleic acid production, also produced high oleic acid at warmer and low humidity sites, especially Bahawalpur. This finding agrees with previous research showing that high temperatures stimulate higher production of oleic acid (Echarte *et al.*, 2010, Akkaya *et al.*, 2019). However, there was no clear-cut advantage of using both parents in breeding programs to develop hybrids with stable high oleic acid (Van Der Merwe *et al.*, 2013, Alberio *et al.*, 2017).

Oleic acid, seed yield and oil content traits are known to be affected by environment. Generally high temperature favored the production of oleic acid (Figure 5B). Evaluation showed that hybrids had higher oleic acid and seed yield at locations with high



Figure 6. Characteristics of newly developed hybrids along with cultivar checks in the spring season. A) Seed yield $(g m^{-2})$; B) oleic acid percentage. Bars showing similar alphabets are statistically insignificant (P>0.05) estimated by least significant difference test.









temperature along with supplementary irrigation during the key reproductive phase. While temperature was optimal for growth at Sargodha, the oleic acid and SY (g m⁻²) were low, possibly due to high humidity and water stress negatively affecting SY due to infestation by fungal diseases (Venkataramanamma *et al.*, 2020). Seed oil contents percentages were favored under mild temperature and

site such as Sargodha had higher oil contents than other sites.

Development of sunflower hybrids with stable high oleic acid, seed yield and oil content was considered an important breeding objective for potential rapid commercialization (Ghaffari *et al.*, 2021). Therefore, this study evaluated sunflower hybrids under diverse environmental conditions and seasons to select hybrids



Figure 8. Relationship between combining ability effects and mean oleic acid contents of inbred lines at four locations, namely, Sargodha (A), Faisalabad (B), Bahawalpur (C) and Multan (D) in Pakistan.



Figure 9. Farm yield trial of high oleic acid sunflower hybrids. Bars showing similar alphabets are statistically insignificant (P≥0.05) estimated by least significant difference test.



The selection of breeding lines with better combining ability could lead to the development of superior highly heterotic hybrids. Generally, lines with higher combining ability values may be utilized in hybrid breeding programs. Combining ability effects showed a high positive relationship with mean oleic acid content at Faisalabad and Bahawalpur (Figure 8), indicating useful gain from selection of this trait. Furthermore, heritability of oleic acid content was medium across locations. This may be due to the response of the high oleic acid mutant genes to environmental factors such as temperature, water availability and humidity (Echarte *et al.*, 2010; Akkaya *et al.*, 2019). Several genomic regions are known to modify oleic acid content in sunflower (Premnath *et al.*, 2016). Inbred lines RH.344, RH.347. RH.345 and C.112. were positive combiners for high oleic acid content and are regarded as promising lines for the development of high oleic acid hybrids.

Conclusions

This study aimed to develop hybrids with high oleic acid contents and SY. Hybrids were evaluated at multiple locations across two seasons. Among the evaluated hybrids, H1, H4, H5 and H10 were the most promising due to their high oleic acid content across the locations during the spring season. Hybrid H10 was the most promising due to its stable and medium oleic acid content (67%) across the locations and between seasons. Hybrid H5 had high oleic acid (79%) and was stable according to the biplot analysis and Kang's ranking. The H5 hybrid also had the highest SY across locations and exhibited higher yield than a low oleic acid standard hybrid. Breeding lines such as RH.344, RH.347. RH.345 and C.112. were positive combiners for high oleic acid content and are regarded as promising lines for the development of high oleic acid hybrids.

References

Akkaya MR, Cil A, Çil AN, Yücel H, Kola O, 2019. The influence of sowing dates on the oil content and fatty acid composition of standard mid-oleic and high-oleic types of sunflower



(Helianthus annuus L.). Food Sci. Tech. 39:448-53.

- Alberio C, Aguirrezábal LA, Izquierdo NG, Reid R, Zuil S, Zambelli A, 2017. Effect of genetic background on the stability of sunflower fatty acid composition in different high oleic mutations. J Sci. Food Agric. 98:4074-84.
- Astrup A, Teicholz N, Magkos F, Bier DM, Brenna JT, King JC, Mente A, Ordovas JM, Volek JS, Yusuf S, Krauss RM, 2021. Dietary saturated fats and health: are the US guidelines evidence-based? Nutrients 13:3305.
- Berville A, Lacombe S, Veillet S, Granier C, Leger S, Jouve P, 2009. Method of selecting sunflower genotypes with high oleic acid content in seed oil. The Patent Cooperation Treaty (PCT) WO 2005/106022 A2.
- Bilgen BB, Daneshvar S, Evci G, Pekcan V, Yilmaz Mİ, Kaya Y, 2018. Determination of high oleic type and broomrape resistant sunflower hybrids by DNA markers. Ekin J. Crop Breed. Genet. 4: 22-30.
- Chernova AI, Gubaev RF, Singh A, Sherbina K, Goryunova SV, Martynova EU, Goryunov DV, Boldyrev SV, Vanyushkina AA, Anikanov NA, Stekolshchikova EA, Yushina EA, Demurin YN, Mukhina ZM, Gavrilova VA, Anisimova IN, Karabitsina YI, Alpatieva NV, Chang PL, Khaitovich P, Mazin PV, Nuzhdin SV, 2021. Genotyping and lipid profiling of 601 cultivated sunflower lines reveals novel genetic determinants of oil fatty acid content. BMC Genomics 22:1-15.
- de Carvalho CGP, Mazzola LF, Mandarino JMG, Dalchiavon FC, 2019. Fatty acid profiles of oil obtained from mid-oleic sunflowers grown in tropical region. J. Am. Oil Chem. Soc. 96: 1019-25.
- Dimitrijević A, Imerovski I, Miladinović D, Cvejić S, Jocić S, Zeremski T, Sakač Z, 2017. Oleic acid variation and markerassisted detection of Pervenets mutation in high-and low-oleic sunflower cross. Crop Breed. Appl. Biotech. 17:235-41.
- Echarte MM, Angeloni P, Jaimes F, Tognetti J, 2010. Night temperature and intercepted solar radiation additively contribute to oleic acid percentage in sunflower oil. Field Crop Res. 119:27-35.
- Eberhardt SA, Russell WA, 1966. Stability parameters for comparing varieties. Crop Sci. 6:36-40.
- Ferreira EB, Cavalcanti PP, Nogueira DA, 2014. ExpDes: an R package for ANOVA and experimental designs. Appl. Math. 5:29-52.
- Finlay KW, Wilkinson GN, 1963. The analysis of adaptation in a plant-breeding programme. Aust. J. Agric. Res. 14:742-54.
- Flagella Z, Rotunno T, Tarantino E, Di Caterina R, De Caro A, 2002. Changes in seed yield and oil fatty acid composition of high oleic sunflower (Helianthus annuus L.) hybrids in relation to the sowing date and the water regime. Eur. J. Agron. 17:221-30.
- Francis, T.R. and Kannenberg, L.W. (1978) Yield stability studies in short-season maize. 1. A descriptive method for grouping genotypes. Canad. J. Plant Sci. 58:1029-34.
- Ghaffari M, Gholizadeh A, Andarkhor SA, Zareei Siahbidi A, Kalantar Ahmadi SA, Shariati F, Rezaeizad A, 2021. Stability and genotype × environment analysis of oil yield of sunflower single cross hybrids in diverse environments of Iran. Euphytica 217:1-11.
- Gupta MK, 2014. Sunflower oil and its applications. Lipid Tech. 26:260-3.
- Joksimović J, Atlagić J, Marinković R, Jovanovic D, 2006. Genetic control of oleic and linoleic acid contents in sunflower. Helia 29:33-40.
- Kang MS, 1988. A rank-sum method for selecting high-yielding, stable corn genotypes. Cereal Res. Comm. 16:113-5.
- Kempthorne O, 1957. An introduction to genetic statistics. New



York: John Wiley and Sons Inc; London: Chapman and Hall Ltd.

- Manalili CJS, Flores EAC, Gaban PBV, Aquino JDC, 2021. Agromorphological characterization and fatty acid composition analysis of selected sunflower accessions. Philippine J. Sci. 150:1255-64.
- Plaisted RL, Peterson LC, 1959. A technique for evaluating the ability of selections to yield consistently in different locations or seasons. Amer. Potato J. 36:381-5.
- Pour-Aboughadareh A, Yousefian M, Moradkhani H, Poczai P, Siddique KHM, 2019. STABILITYSOFT: a new online program to calculate parametric and non-parametric stability statistics for crop traits. Appl. Plant Sci. 7:e01211.
- Premnath A, Narayana M, Ramakrishnan C, Kuppussamy S, 2016. Mapping quantitative trait loci controlling oil content oleic acid and linoleic acid content in sunflower (Helianthus annuus L). Mol. Breed. 36:106.
- Qadir G, Ahmad S, Ul-Hassan F, Cheema M, 2006. Oil and fatty acid accumulation in sunflower as influenced by temperature variation Pak. J. Bot. 38:1137-47.
- Rauf S, 2019. Breeding strategies for sunflower (Helianthus annuus L.) genetic improvement. In: Al-Khayri J, Jain S, Johnson D (eds). Advances in Plant Breeding Strategies: Industrial and Food Crops. Cham, Springer. pp. 637-73.
- Rauf S, Jamil N, Tariq SA, Khan M, Kausar M, Kaya Y, 2017. Progress in modification of sunflower oil to expand its industrial value. J. Sci. Food Agric. 97:1997-2006.
- Rauf S, Ortiz R, Shehzad M, Haider W, Ahmed I, 2020. The exploitation of sunflower (Helianthus annuus L.) seed and

other parts for human nutrition, medicine and the industry. Helia 43:167-84.

- Reynolds SG, 1970. The gravimetric method of soil moisture determination Part IA study of equipment, and methodological problems. J. Hydrol. 11:258-73.
- Rogers SO, Bendich AJ, 1985. Extraction of DNA from milligram amounts of fresh, herbarium and mummified plant tissues. Plant Mol. Biol. 5:69-76.
- Romano R, Filosa G, Pizzolongo F, Durazzo A, Lucarini M, Severino P Souto EB, Santini A, 2021. Oxidative stability of high oleic sunflower oil during deep-frying process of purple potato Purple Majesty. Heliyon 7:e06294.
- Shukla GK, 1972. Genotype stability analysis and its application to potato regional trials. Crop Sci. 11:184-90.
- Van Der Merwe R, Labuschagne M, Herselman L, Hugo A, 2013. Stability of seed oil quality traits in high and mid-oleic acid sunflower hybrids. Euphytica 193:157-68.
- Venkataramanamma K, Rajendran L, Gopalakrishnan C, 2020. Sunflower (Helianthus annuus L.) diseases and their management by integrated approach. In: Srivastava JN, Singh AK (eds). Diseases of Field Crops: Diagnosis and Management. Palm Bay, FL: Apple Academic Press. pp. 287-306.
- Wricke G, 1962. Evaluation method for recording ecological differences in field trials. Z Pflanzenzücht, 47:92-6.
- Yan W, Kang MS, 2002. GGE biplot analysis: a graphical tool for breeders geneticists and agronomists Boca Raton, FL: CRC press.
- Zambelli A, 2021. Current status of high oleic seed oils in food processing. J. Am. Oil Chem. Soc. 98:129-37.