Qualitative characterisation of cultivated and wild edible plants: Mineral elements, phenols content and antioxidant capacity

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Abstract

This study investigated the qualitative characteristics of several edible wild herbaceous species, including those most consumed in Foggia Province (southern Italy). Analysis of qualitative characteristics was performed for the edible parts of 11 wild species (Beta vulgaris L., Foeniculum vulgare Miller, Centaurea solstitialis L., Cichorium intybus L., Scolymus hispanicus L., Sonchus oleraceus L., Borago officinalis L., Erucastrum sativum L., Zea mays L., and Diplotaxis tenuifolia L.) and three cultivated species (C. intybus, B. officinalis, D. tenuifolia). The plants were collected from areas in the Foggia countryside, and the edible part of each species was analysed for dry matter, protein, cation and anion contents as well as total phenols and antioxidant activities. Among the cations, calcium was the most differentiated among species, ranging 784 mg kg–1 fresh weight (Fw) for B. vulgaris to 5886 mg kg–1 Fw for S. hispanicus. The nitrate contents were also highly variable, from 75 mg kg –1 Fw for C. intybus to 586 mg kg –1 Fw for D. tenuifolia. Total polyphenols ranged from 1054 mg gallic acid equivalents (GAE) mg kg–1 Fw for C. solstitialis to 3664 mg GAE mg kg–1 Fw for S. arvensis. Antioxidant activities ranged from 839 mg Trolox equivalent (TE) mg kg–1 Fw for B. vulgaris to 5658 mg TE kg–1 Fw for C. intybus. Significant differences were also noted between wild and cultivated plants in the qualitative parameters. Total polyphenols and antioxidant activity were higher in wild C. intybus and B. officinalis than in their cultivated counterparts. Multivariate analysis (cluster analysis and linear discriminant analysis) allowed integration of the ANOVA data to determine the qualitative characteristics of the wild species that contribute most to group differences. The results of the present study aims to improve current knowledge about edible wild species as vegetable sources in the Mediterranean diet.

Introduction

Wild edible plants represent a particular aspect of the genetic biodiversity of vegetables and cultivated species as ecotypes and local varieties (Tarantino et al., 2011), and they are also an important food source, especially in the Mediterranean Basin (Boari et al., 2013; Hadjichambis et al., 2008). In this region, the environment is characterised by a great wealth of endemic flora, the diversity of which led to their use in the past as food for humans and animals as well as in folklore, veterinary medicine, cosmetics, food flavouring, beverages, energy production, crafts, and gardening (Marzi and Tedone, 2009).

In Europe, Italy is the country with the greatest plant biodiversity and hosts 6711 plant species (Conti et al., 2005). Several studies carried out over the last few decades have estimated that approximately 700 plant species belonging to 93 botanical families are used as vegetables or for seasoning (Bianco, 1989, 1990, 1997; Bianco and Maackackova, 2002). The use of wild edible plants for food is a common practice in Italy (Aliotta, 1987, 1993, 2013; Hadjichambis et al., 2008). In this region, the environment is characterised by a great wealth of endemic flora, the diversity of which led to their use in the past as food for humans and animals as well as in folklore, veterinary medicine, cosmetics, food flavouring, beverages, energy production, crafts, and gardening (Marzi and Tedone, 2009).

According to 3000 recipes from the southern regions of Italy, there were ancient dishes that incorporated wild edible plants, many of which were (and still are) used to prepare soups and stews, herb omelettes, and salads. However, the compositions of the dishes are very variable in terms of the number of species used; e.g., for a mixed soup, approximately 40 species were used to give the soup a balanced flavour by making it sweeter, more bitter and/or spicy (Marzi and Tedone, 2009).

Foggia is the northernmost Province of Apulia and is characterised by plains, valleys, hills, and mountains that rise up to 1151 m above sea level, and there are natural and semi-natural areas, uncultivated arable land, large areas with only sheep tracks, and banks along streams and gullies. Plants grow in all of these areas without the use of chemicals, and it is still possible to collect many...
species of edible wild plants, thus further promoting their traditional culinary uses. The knowledge of the use of these plants is deep-rooted in Foggia province, where the older people are the connoisseurs and collectors of wild herbs and pass down this knowledge from father to son and mother to daughter. Many Italian people share a passion for food plants, and in Foggia province this is enriched by the produce in the local markets that constantly satisfy the needs of consumers.

The therapeutic and nutritional properties of wild edible plants have been the subject of numerous studies (Vitalini et al., 2006; Pardo de Santayana et al., 2007), and these plants are generally characterised by high nutritional but low energy values (Renna, 2017). Compared to their corresponding cultivated species, wild food plants have higher fibre content (Leonti et al., 2006), are richer in vitamins, minerals, polyphenols, antioxidants, and flavonoids (Pieroni et al., 2002), and have very low lipid levels (Trichopoulou et al., 2000). Natural products with antioxidant activity can enhance the endogenous defences of organisms against exposure to free radicals, and epidemiological evidence suggests that consuming a high amount of vegetables that are rich in phenolic compounds and that have high antioxidant activities in the diet is associated with a reduced incidence of coronary heart disease and some cancers (Cook and Samman, 1996; Peterson and Dwyner, 1998).

In this context, wild edible plants can have important beneficial effects in the prevention of several modern chronic diseases, such as age- and heart-related pathologies, diabetes and cancers (Finkel et al., 2000). Natural products with antioxidant activity can enhance the endogenous defences of organisms against exposure to free radicals, and epidemiological evidence suggests that consuming a high amount of vegetables that are rich in phenolic compounds and that have high antioxidant activities in the diet is associated with a reduced incidence of coronary heart disease and some cancers (Cook and Samman, 1996; Peterson and Dwyner, 1998).

Wild, naturally growing vegetables are considered healthier than cultivated species as they should be free of chemicals, such as residues from the fertilisers and pesticides. However, if they are collected along roads with heavy vehicular traffic, they might contain toxic substances, including heavy metals (Alloway, 2009). Wild, naturally growing vegetables are considered healthier than cultivated species as they should be free of chemicals, such as residues from the fertilisers and pesticides. However, if they are collected along roads with heavy vehicular traffic, they might contain toxic substances, including heavy metals (Alloway, 2009).

More important than the chemical composition of wild plants is the content in health-promoting secondary compounds, such as flavonoids, phenolic acids, and their derivatives (Lamuela-Raventós et al., 2003). These compounds have antioxidant properties and may prevent chronic diseases associated with oxidative stress (Santamaria, 2006; Guil Guerrero et al., 2004; Clark et al., 2006; Renna et al., 2014). Moreover, high nitrate contents represent a serious threat to human health, and if they are collected along roads with heavy vehicular traffic, they might contain toxic substances, including heavy metals (Alloway, 2009). Wild, naturally growing vegetables are considered healthier than cultivated species as they should be free of chemicals, such as residues from the fertilisers and pesticides. However, if they are collected along roads with heavy vehicular traffic, they might contain toxic substances, including heavy metals (Alloway, 2009).

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The study plants were collected from autumn of 2015 until spring of 2016 in the Foggia countryside (Figure 1) when these wild species are most suitable for consumption. Wild species were sampled more than 200 metres from roads (buffer zone) to avoid possible contamination. Forty plants of each species were manually collected and pooled to form a single sample. The cultivated plant species, which were grown in open field according to local agronomic standards, were collected from some Apulian farms. For these species the soil was lowing 30 cm depth, and before transplanting or sowing, its surface was milked. Pretransplanting/sowing fertilisation was applied to the soil by distributing 60 kg ha⁻¹ N and 30 kg ha⁻¹ P₂O₅. C. Intybus was trans-

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**Materials and methods**

**Agronomic conditions and sampling procedure**

This study was conducted at the Department of Agriculture, Food and Environment of the University of Foggia, and it qualitatively analysed 11 wild edible plants that are deeply rooted in the traditional cuisine of Foggia Province: *Beta vulgaris*, *Foeniculum vulgare* Miller, *Centaurea solstitialis*, *Cichorium intybus*, *Scolymus hispanicus*, *Sonchus oleraceus*, *Borago officinalis*, *Diplotaxis erucoides*, *Diplotaxis tenuifolia* (L.) DC, *Sinapis arvensis*, and *Portulaca oleracea*. In addition, cultivated plants of *C. intybus*, *B. officinalis*, and *D. tenuifolia* were collected. Table 1 provides the details of each plant species including botanical family, scientific name, common English name, harvesting time, edible parts, and culinary uses.

Through the year; best in spring

Leaf, flower, seed

Leaves in Spring, stems and seeds in Summer-Autumn

Leaves, root

Leaves, flower

Leaf, flower

Leaf, seed, flower

Leaf, flower

Leaf

Leaf

Leaf, flower

Leaf, flower

Leaf

Leaf, flower

Leaves, root

Leaves, flower

Leaves, flower

Leaves, flower

Leaves, flower

Table 1. Details of the wild herbaceous edible plants in this study.

<table>
<thead>
<tr>
<th>Family</th>
<th>Scientific name</th>
<th>CEN</th>
<th>Harvesting time</th>
<th>Edible part</th>
<th>Culinary use</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amaranthaceae</td>
<td><em>Beta vulgaris</em> L.</td>
<td>Wild beet</td>
<td>October to April</td>
<td>Leaf, stem</td>
<td>Boiled, salted rustic cakes, soups</td>
</tr>
<tr>
<td>Apiaceae</td>
<td><em>Foeniculum vulgare</em> M.</td>
<td>Wild fennel</td>
<td>Leaves in Spring, stems and seeds in Summer-Autumn</td>
<td>Leaf, fresh seed, dry seed</td>
<td>Boiled, seasoning, salads, pickles or pickled, soups</td>
</tr>
<tr>
<td>Asteraceae</td>
<td><em>Centaurea solstitialis</em></td>
<td>Yellow cornflower</td>
<td>April to June</td>
<td>Leaf, root</td>
<td>Boiled, salads, rustic pies, soups</td>
</tr>
<tr>
<td></td>
<td><em>Cichorium intybus</em> L.</td>
<td>Wild chicory</td>
<td>October to April</td>
<td>Root, flower, leaf</td>
<td>Boiled, soups</td>
</tr>
<tr>
<td></td>
<td><em>Scolymus hispanicus</em></td>
<td>Common cardogna</td>
<td>December to May</td>
<td>Flower, rib leaf</td>
<td>Boiled, soups</td>
</tr>
<tr>
<td></td>
<td><em>Sonchus oleraceus</em> L.</td>
<td>Sow thistles</td>
<td>Throughout the year; best in spring</td>
<td>Leaf, seed, flower</td>
<td>Boiled, soups</td>
</tr>
<tr>
<td>Boraginaceae</td>
<td><em>Borago officinalis</em> L.*</td>
<td>Starflower</td>
<td>October to June</td>
<td>Leaf, flower</td>
<td>Boiled, omelettes, salads, salted, rustic cakes, soups</td>
</tr>
<tr>
<td>Brassicaceae</td>
<td><em>Diplotaxis erucoides</em> (L.) DC</td>
<td>Purplish rocket</td>
<td>September to March</td>
<td>Leaf</td>
<td>Boiled, salads, rustic pies, soups</td>
</tr>
<tr>
<td></td>
<td><em>Diplotaxis tenuifolia</em> L.*</td>
<td>Wild rocket</td>
<td>Throughout the year</td>
<td>Leaf</td>
<td>Boiled, salads, seasoning, savoury or rustic cakes</td>
</tr>
<tr>
<td></td>
<td><em>Sinapis arvensis</em> L.</td>
<td>Wild mustard</td>
<td>March to June</td>
<td>Leaf</td>
<td>Boiled, sauces, seasoning, savoury or rustic cakes</td>
</tr>
<tr>
<td>Portulacaceae</td>
<td><em>Portulaca oleracea</em> L.</td>
<td>Purslane</td>
<td>June to September</td>
<td>Leaf</td>
<td>Boiled, salads, omelettes, pickles or pickled, and soups</td>
</tr>
</tbody>
</table>

CEN, common English name. *Both wild and cultivated plants of these species were analysed*.
planted on October 8th, 2015 (10 plant m$^{-2}$) and harvested on February 22nd, 2016; *B. Officinalis* was directly sown in the field on April 10th 2016 (16.7 plant m$^{-2}$) and harvested on June 20th, 2016; *D.tenuifolia* was sown on April 5th, 2016 (66.7 plant m$^{-2}$) and harvested on June 30th, 2016. For all cultivated species the pest and weed control were performed according to local management practices. Drip irrigation system was used for their irrigation. This method comprised a single pipe, with drippers at a 2 L h$^{-1}$ flow rate. The amount of irrigation water applied to *C. Intybus*, *B. Officinalis* and *D.tenuifolia* during the whole crop cycles was 1400, 1500 and 1700 m$^{3}$ ha$^{-1}$, respectively.

All sample plants were transported to the laboratory under refrigerated conditions for analysis. Fifty-six samples (four replicates of 11 wild and three cultivated species) were gently cleaned and separated into edible and waste parts, which included the older leaves and stems that are normally removed during preparation for cooking.

### Qualitative analysis

For each sample, four replicates of the edible parts were analysed for dry matter, protein, cations and anions. Total polyphenols and antioxidant activity were also determined.

To measure dry matter, samples of the edible fresh portions of the plants were dried to a constant weight in a forced-air oven at 65°C, and these data are expressed as g per 100 g fresh weight (Fw). Total nitrogen was determined using a LECO CHN-600 determinator, and protein content was calculated by multiplying the total nitrogen by 6.25 and is expressed as g per 100 g Fw.

The anion (fluoride, chloride, nitrate, phosphate and sulfate) and cation (sodium, potassium, magnesium, calcium) contents were determined using ion-exchange chromatography (ICS-1100; Dionex Corporation, Sunnyvale, CA, USA). The anions were extracted from 0.5-g dried and ground samples with 50 mL 3.5 nmol L$^{-1}$ NaCO$_3$ and 1.0 mmol L$^{-1}$ NaHCO$_3$, and the extracts were analysed using a guard column and an analytical column (Ionpac AG14 and AS14, respectively). The data are expressed as mg kg$^{-1}$ Fw. For the cations, 1.0-g dried and ground samples were used with the ash produced in a muffle furnace at 550 °C that was then digested in 20 mL 1.0 mol L$^{-1}$ HCl in boiling water (99.5±0.5°C) for 30 min. The resulting solution was filtered, diluted, and analysed using a guard column and analytical column (Ionpac CG12A and CS12A, respectively). The data are expressed as mg kg$^{-1}$ Fw (Renna et al., 2014).

The total phenol content was determined according to the method of Singleton and Rossi (1965) and expressed as mg gallic acid equivalents (GAE) per kg Fw. The antioxidant activity of the plant extracts was determined by the method of Brand-Williams et al. (1995) with modifications. The diluted samples (50 μL) were added to 950 μL 1,1-diphenyl-2-picrylhydrazyl (DPPH) solution to initiate the reactions, and after incubation overnight at 23°C, the absorbance was read using a spectrophotometer (Perkin Elmer UV/VIS LAMBDA 45) at 515 nm. Trolox was used as the standard for analysis, and the antioxidant activity is reported as mg Trolox equivalents (TE) per kg Fw.

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Figure 1. Map of Italy showing the location of Foggia Province.
Statistical analysis

The datasets were analysed according to the basic assumptions for analysis of variance (ANOVA). The normality of the distributions and the common variance of the experimental error were verified by Shapiro–Wilk and Bartlett’s tests, respectively. When required, Box–Cox transformations (Box and Cox, 1964) were applied prior to analysis, and for all data, the ANOVA was performed according to a completely randomised design with four replicates. The differences between the means were determined by Tukey’s honest significance difference post hoc tests at the 5% probability level, and bivariate statistical methods were applied (linear regression analysis) to determine the relationships between the different qualitative characteristics of the herbaceous food plants.

Furthermore, the qualitative parameters of the wild edible plants were jointly considered using a multivariate approach. Cluster analysis (Everitt, 1980; Aldenderfer and Blashfield, 1984) was used to find truly homogeneous groups of species. Ward’s minimum variance hierarchical clustering was performed using an agglomerative approach and Ward’s linkage, and at each cluster definition, samples were added into superior clusters to minimise the within-cluster sum of squares or maximise the between-cluster sum of squares (Podani, 2007; Moore et al., 2010). To compare differences among clusters, ANOVA and Tukey’s tests were used for all continuous variables (5% probability level).

Linear discriminant analysis (Moore et al., 2010; McLaclan, 1992; Webb, 2002; Cammareri et al., 2004) was used to best explain the differences between the different cluster groups obtained by cluster analysis and to evaluate which of the main parameters play the most important roles in determining these differences. The independent variables defined 13 qualitative parameters, and the dependent variable was the wild species. The first two linear discriminant canonicals (CNs) accounted for the largest portion of the variability in the data and were considered for data interpretation. The selected CNs were graphically represented in biplots, in which the discriminant scores were plotted as symbols, and the standardised coefficients for each of the original variables were reported as vectors and used to interpret the data. Before the cluster and linear discriminant analyses, the values of each parameter were correctly standardised.

The ANOVA, cluster analysis and linear discriminant analysis were performed using the JMP software package, version 8.1 (SAS Institute Inc., Cary, NC, USA), and graphs were constructed using SigmaPlot (Systat Software, Chicago, IL, USA).

Results

Qualitative characteristics of wild herbaceous food plants

Figures 2-5 show the means, standard errors, and significance of the analysed qualitative parameters for the edible parts of each wild edible plant species. All of the considered parameters differed significantly among the species.

Dry matter content in the tested plants ranged from 7.1 g 100 g–1 Fw for B. officinalis to 16.2 g 100 g–1 Fw for D. tenuifolia. This highest mean value was not significantly different from the means for F. vulgaris and C. solstitialis. The values for these species were followed by those for S. arvensis (12.3 mg per 100 g Fw) and D. erucoides (10.4 mg per 100 g Fw), and the means were significantly lower for all the other species (Figure 2A).

The mean protein content ranged from 1.9 mg 100 g–1 Fw for C. intybus to 4.8 mg 100 g–1 Fw for D. tenuifolia, but this high value was not significantly different from the protein content of F. vulgaris, S. arvensis, D. erucoides, and C. solstitialis. These wild species appeared to have very high protein levels with respect to common vegetables (lettuce, spinach, parsley, carrots and cabbage) (McCullom, 1992; Singh et al., 2001). All the other species showed low values (never less than 2 mg 100 g–1 Fw) that were not significantly different from each other (Figure 2B).

The mean values of the minerals (i.e., sodium, potassium, magnesium and calcium) that are important for human nutrition are reported in Figure 3; generally, the differences between the minimum and maximum values for the analysed species were approximately 3-fold for potassium and approximately 7-8-fold for magnesium, calcium, and sodium. The highest sodium content was 2992 mg kg–1 Fw for B. vulgaris, which was significantly higher than that for all the other species for which the means ranged from 357 mg kg–1 Fw for D. erucoides to 802 mg kg–1 Fw for D. tenuifolia but were not significantly different (Figure 3A).

The potassium levels ranged from 1870 mg kg–1 Fw for P. oleracea to 6090 mg kg–1 Fw for S. hispanicus, which showed a value
similar to that of *D. tenuifolia* and *B. officinalis* followed by *F. vulgaris* and *C. intybus*. All the other species showed significantly lower potassium levels (<4000 mg kg⁻¹ Fw) with no significant differences among them (Figure 3B).

The magnesium levels ranged from 79 mg kg⁻¹ Fw for *B. vulgaris* to 646 mg kg⁻¹ Fw for *S. hispanicus*, but the high magnesium value for *S. hispanicus* was not significantly different from that of *D. tenuifolia*, which was not significantly different from that of *C. solstitialis*. All the other species showed lower magnesium contents from 259 to 385 mg kg⁻¹ Fw, with no significant differences among them (Figure 3C).

Calcium levels were highest for *S. hispanicus* (5886 mg kg⁻¹ Fw) and significantly differed from those of all the other species. The lowest calcium content was 784 mg kg⁻¹ Fw for *B. vulgaris*, although this value was not significantly different from that of *P. oleracea*, *S. oleraceus*, *B. officinalis*, *C. intybus*, and cultivated *B. officinalis*. The other species showed intermediate calcium levels from 2550 to 3750 mg kg⁻¹ Fw (Figure 3D). The differences between the minimum and maximum anion contents (i.e., nitrates, fluorides, chlorides, phosphorus and sulfur; Figure 4A-C) of these different species ranged from approximately 11-12-fold for fluorides, chlorides and sulfur to approximately 50-fold for nitrates to the greatest difference of approximately 137-fold for phosphorus.

The nitrate contents ranged from 75 mg kg⁻¹ Fw for wild *C. intybus* to 3874 mg kg⁻¹ Fw for *D. tenuifolia*, which was not significantly different from that of *C. solstitialis* (3579 mg kg⁻¹ Fw) that, in turn, was not significantly different from that of *S. arvensis* (3028 mg kg⁻¹ Fw) followed by *D. erucoides* (2024 mg kg⁻¹ Fw).

The highest fluoride levels were found in *D. tenuifolia* (180 mg kg⁻¹ Fw), which differed significantly from all the other species. The values ranged from 16 mg kg⁻¹ Fw for *P. oleracea* to 126 mg kg⁻¹ Fw for *C. intybus*, which was not significantly different from *F. vulgaris*. The other species showed low fluoride levels ranging from 16 to 69 mg kg⁻¹ Fw.

The highest level of chlorides was 9348 mg kg⁻¹ Fw for *C. solstitialis*, which differed significantly from all of the other species, whose values ranged from 832 mg kg⁻¹ Fw for *S. arvensis* to 3070 mg kg⁻¹ Fw for *F. vulgaris*. In turn, the value of the latter was not significantly different from that of *S. hispanicus* followed by all of the other species whose lower values were not significantly different.

The phosphorus levels were significantly higher in *C. intybus* (511 mg kg⁻¹ Fw) although they were not significantly different from those of *S. arvensis* (437 mg kg⁻¹ Fw) and *C. solstitialis* (406 mg kg⁻¹ Fw). All the other species showed levels ranging from 4 mg kg⁻¹ Fw for *F. vulgaris* to 179 mg kg⁻¹ Fw for *D. tenuifolia*.

Finally, the highest sulfur level of 741 mg kg⁻¹ Fw was seen in *S. arvensis*, although this value was not significantly different from...
645 mg kg\(^{-1}\) Fw for \(P.\ oleracea\) and 616 mg kg\(^{-1}\) Fw for \(F.\ vulgare\). The levels of these two species were not significantly different from the value of 543 mg kg\(^{-1}\) Fw for \(D.\ tenuifolia\), which was followed by the intermediate levels of 399.3 mg kg\(^{-1}\) Fw for \(S.\ hispanicus\) and 358 mg kg\(^{-1}\) Fw for \(C.\ intybus\).

The total polyphenol content of the analysed plants ranged from 1054 mg GAE kg\(^{-1}\) Fw for \(C.\ solstitialis\) to 3664 mg GAE kg\(^{-1}\) Fw for \(S.\ arvensis\), which was not significantly different from the total polyphenols for wild \(C.\ intybus\) (3073 mg GAE kg\(^{-1}\) Fw), \(B.\ officinalis\) (2728 mg GAE kg\(^{-1}\) Fw) and \(S.\ hispanicus\) (2586 mg GAE kg\(^{-1}\) Fw). The value for this last species was followed by those for \(S.\ oleraceus\) and \(D.\ erucoides\). The low total polyphenol contents for the remaining species ranged from 1054 to 1639 mg GAE kg\(^{-1}\) Fw (Figure 5A).

There were large variations in antioxidant activities that ranged from 839 mg TE kg\(^{-1}\) Fw for \(B.\ vulgaris\) to 5658 mg TE kg\(^{-1}\) Fw for \(C.\ intybus\), whose value was significantly different from those of the other species. This activity level was followed by those of \(B.\ officinalis\) (3928 mg TE kg\(^{-1}\) Fw), \(D.\ erucoides\) (3920 mg TE kg\(^{-1}\) Fw), \(S.\ arvensis\) (3669 mg TE kg\(^{-1}\) Fw), and \(S.\ hispanicus\) (2719 mg TE kg\(^{-1}\) Fw), which were not significantly different from each other, and low values of <2151 mg TE kg\(^{-1}\) Fw were observed for the other species (Figure 5B).

**Qualitative characteristics of wild vs cultivated herbaceous edible plants**

The qualitative characteristics of three wild and cultivated species, \(C.\ intybus\), \(B.\ officinalis\), and \(D.\ tenuifolia\), were also compared. Table 2 shows the means, standard errors, and significance values of these parameters for the edible parts of each species.

Dry matter contents of these three species did not differ between the wild and cultivated plants, and the same was observed for the protein content.

Sodium levels were significantly higher in cultivated \(C.\ intybus\) (902 mg kg\(^{-1}\) Fw) compared to wild \(C.\ intybus\) (720.2 mg kg\(^{-1}\) Fw). In contrast, wild \(D.\ tenuifolia\) showed significantly higher sodium levels than cultivated \(D.\ tenuifolia\), and the same pattern was seen for potassium levels. No differences in potassium levels were observed between the wild and cultivated \(C.\ intybus\) and \(B.\ officinalis\) plants.

The magnesium and calcium levels also showed no differences between these three wild and cultivated species. The fluoride content did not differ between the wild and cultivated \(C.\ intybus\) and \(B.\ officinalis\), while they were significantly higher in wild (180 mg kg\(^{-1}\) Fw) than in cultivated \(D.\ tenuifolia\) (117 mg kg\(^{-1}\) Fw). The chloride levels were significantly higher in cultivated \(C.\ intybus\) (3740 mg kg\(^{-1}\) Fw) than wild plants (1807 mg kg\(^{-1}\) Fw), whereas no differences were observed between both wild and cultivated \(B.\ officinalis\) and \(D.\ tenuifolia\). The phosphorus levels in \(C.\ intybus\) and \(B.\ officinalis\) did not differ between wild and cultivated plants, but cultivated \(D.\ tenuifolia\) showed significantly higher phosphorus levels (562.5 mg kg\(^{-1}\) Fw) than the wild plants (178.7 mg kg\(^{-1}\) Fw). The same was observed for sulfur levels, which did not differ between wild and cultivated \(C.\ intybus\) and \(B.\ officinalis\), while significantly higher levels were seen in cultivated \(D.\ tenuifolia\) (863.5 mg kg\(^{-1}\) Fw) relative to wild plants (741.5 mg kg\(^{-1}\) Fw).

The nitrate contents of the wild \(C.\ intybus\), \(B.\ officinalis\) and \(D.\ tenuifolia\) were significantly lower (75.5; 1060.7 and 3873.9 mg kg\(^{-1}\) Fw, respectively) compared to those of cultivated plants (2191.8; 1438 and 4853.5 mg kg\(^{-1}\) Fw, respectively).

Only considering these three species, the total polyphenols and antioxidant activity were significantly highest in \(C.\ intybus\) (2688 mg GAE kg\(^{-1}\) Fw)
mg GAE kg⁻¹ Fw and 4111 mg TE g⁻¹ Fw, respectively) followed by *B. officinalis* (2333 mg GAE kg⁻¹ Fw and 3141 mg TE kg⁻¹ Fw, respectively) and *D. tenuifolia* (1558 mg GAE kg⁻¹ Fw and 1357 mg TE kg⁻¹ Fw, respectively). Moreover, the total polyphenols and antioxidant activities were significantly higher in the wild plants compared to the cultivated plants for both *C. intybus* and *B. officinalis*, but there were no differences between wild and cultivated *D. tenuifolia* (Table 2). Finally, there was a positive and highly significant linear correlation between the total polyphenol contents and antioxidant activities of all of the analysed species (Figure 6).

**Cluster and linear discriminant analysis results**

The cluster analysis of all the qualitative characteristics of the 11 herbaceous species identified five clusters (Figure 7). The best number of clusters was defined through the inverse relationship between the number of clusters and the mean distances among them. Thus, an increase in the number of clusters did not result in any substantial reduction in the distances between the clusters. As a result of this analysis, the mean of each qualitative parameter (±standard error) as well as the significance level (P-value) of the ANOVA for each variable were represented by each cluster (Table 3). Significant differences among the variables were based on P<0.05.

Cluster 1 comprises *B. vulgaris* and *P. oleracea* and is characterised by very low values for all the qualitative parameters considered except for the sodium level. Cluster 2 comprises *C. intybus*, *S. hispanicus* and *S. arvensis*, all of which show high antioxidant activity and total phenol, potassium and calcium contents. Cluster 3 comprises the two wild species of *F. vulgaris* and *D. tenuifolia*, and it is distinguished by higher dry matter, fluoride, potassium, and sulfate contents than the other clusters. Cluster 4 contains a single species, *C. solstitialis*, and it is primarily characterised by

### Table 2. Comparisons between the three analysed wild and cultivated species.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Units</th>
<th>B. officinalis L.</th>
<th>Wild</th>
<th>Cultivated</th>
<th>P</th>
<th>B. officinalis L.</th>
<th>Wild</th>
<th>Cultivated</th>
<th>P</th>
<th>D. tenuifolia L.</th>
<th>Wild</th>
<th>Cultivated</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dry matter</td>
<td>mg 100 g⁻¹ Fw</td>
<td>9.45±0.70</td>
<td>14.0±0.12</td>
<td>ns</td>
<td>6.18±0.15</td>
<td>8.13±0.38</td>
<td>ns</td>
<td>16.20±0.51</td>
<td>9.68±0.29</td>
<td>ns</td>
<td></td>
<td></td>
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<tr>
<td>Protein</td>
<td>mg 100 g⁻¹ Fw</td>
<td>1.88±0.22</td>
<td>2.94±0.28</td>
<td>ns</td>
<td>1.30±0.10</td>
<td>2.27±0.07</td>
<td>ns</td>
<td>4.78±0.14</td>
<td>2.75±0.10</td>
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<td></td>
</tr>
<tr>
<td>Sodium</td>
<td>mg kg⁻¹ Fw</td>
<td>720.2±15.8</td>
<td>901.8±52.4</td>
<td>*</td>
<td>637.1±45.8</td>
<td>524.0±51.3</td>
<td>ns</td>
<td>802.3±19.4</td>
<td>282.5±25.6</td>
<td>*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Potassium</td>
<td>mg kg⁻¹ Fw</td>
<td>4264.2±39.6</td>
<td>4989.2±89.5</td>
<td>ns</td>
<td>5219.4±489.7</td>
<td>4385.5±550.3</td>
<td>ns</td>
<td>5934.9±82.5</td>
<td>2643.4±327.1</td>
<td>*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Magnesium</td>
<td>mg kg⁻¹ Fw</td>
<td>395.5±11.6</td>
<td>439.5±28.0</td>
<td>ns</td>
<td>383.5±20.4</td>
<td>295.5±34.3</td>
<td>ns</td>
<td>514.0±10.8</td>
<td>428.8±33.6</td>
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<td></td>
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<td></td>
</tr>
<tr>
<td>Calcium</td>
<td>mg kg⁻¹ Fw</td>
<td>2160.6±296.9</td>
<td>2671.8±226.5</td>
<td>ns</td>
<td>1966.6±76.3</td>
<td>2424.8±262.4</td>
<td>ns</td>
<td>2351.0±40.1</td>
<td>2789.8±118.7</td>
<td>ns</td>
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<td></td>
</tr>
<tr>
<td>Fluorides</td>
<td>mg kg⁻¹ Fw</td>
<td>125.8±20.8</td>
<td>76.1±7.2</td>
<td>ns</td>
<td>24.5±4.1</td>
<td>41.3±7.7</td>
<td>ns</td>
<td>180.1±5.9</td>
<td>117.5±13.8</td>
<td>*</td>
<td></td>
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<tr>
<td>Chlorides</td>
<td>mg kg⁻¹ Fw</td>
<td>1807.2±182.8</td>
<td>3740.3±377.0</td>
<td>*</td>
<td>1067.0±71.3</td>
<td>2021.3±234.2</td>
<td>ns</td>
<td>1682.0±38.1</td>
<td>1358.0±97.5</td>
<td>ns</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nitrites</td>
<td>mg kg⁻¹ Fw</td>
<td>75.5±14.7</td>
<td>2191.8±184.4</td>
<td>*</td>
<td>1006.7±137.9</td>
<td>1438.3±108.6</td>
<td>*</td>
<td>3873.9±68.0</td>
<td>4853.5±119.2</td>
<td>*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phosphorus</td>
<td>mg kg⁻¹ Fw</td>
<td>511.2±31.5</td>
<td>498.5±63.2</td>
<td>ns</td>
<td>131.8±14.4</td>
<td>121.5±5.2</td>
<td>ns</td>
<td>178.7±10.9</td>
<td>562.5±56.2</td>
<td>*</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sulfur</td>
<td>mg kg⁻¹ Fw</td>
<td>399.3±35.1</td>
<td>358.3±41.8</td>
<td>ns</td>
<td>176.3±17.3</td>
<td>186.5±17.0</td>
<td>ns</td>
<td>741.5±6.4</td>
<td>863.5±21.9</td>
<td>*</td>
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</tr>
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</table>

### Table 3. Results of the analysis of variance for all qualitative parameters for the different clusters.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Units</th>
<th>Cluster 1 (n=8)</th>
<th>Cluster 2 (n=12)</th>
<th>Cluster 3 (n=8)</th>
<th>Cluster 4 (n=12)</th>
<th>Cluster 5 (n=3)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dry matter</td>
<td>mg 100 g⁻¹ Fw</td>
<td>7.94±0.37</td>
<td>9.86±0.65</td>
<td>15.59±0.40</td>
<td>14.43±0.22</td>
<td>8.27±1.12</td>
</tr>
<tr>
<td>Protein</td>
<td>mg 100 g⁻¹ Fw</td>
<td>2.2±0.11</td>
<td>2.69±0.34</td>
<td>4.77±0.53</td>
<td>3.11±0.13</td>
<td>2.57±0.40</td>
</tr>
<tr>
<td>Sodium</td>
<td>mg kg⁻¹ Fw</td>
<td>1705.1±496.0</td>
<td>839.0±243.4</td>
<td>681.3±54.9</td>
<td>621.3±5.9</td>
<td>494.4±8.5</td>
</tr>
<tr>
<td>Potassium</td>
<td>mg kg⁻¹ Fw</td>
<td>2395.7±241.6</td>
<td>4676.3±327.1</td>
<td>5186.5±100.2</td>
<td>2610.0±101.9</td>
<td>4198.2±550.0</td>
</tr>
<tr>
<td>Magnesium</td>
<td>mg kg⁻¹ Fw</td>
<td>169.1±57.2</td>
<td>459.9±46.7</td>
<td>462.3±34.6</td>
<td>482.3±7.5</td>
<td>354.2±34.9</td>
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<tr>
<td>Calcium</td>
<td>mg kg⁻¹ Fw</td>
<td>1034.5±88.6</td>
<td>3867.9±542.3</td>
<td>3150.5±339.4</td>
<td>2758.0±113.3</td>
<td>2150.5±521.8</td>
</tr>
<tr>
<td>Fluorides</td>
<td>mg kg⁻¹ Fw</td>
<td>23.2±5.0</td>
<td>85.0±11.3</td>
<td>140.6±16.5</td>
<td>23.0±8.8</td>
<td>31.8±11.6</td>
</tr>
<tr>
<td>Chlorides</td>
<td>mg kg⁻¹ Fw</td>
<td>1059.6±64.6</td>
<td>1585.4±184.7</td>
<td>2376.1±286.4</td>
<td>9348.0±361.8</td>
<td>1549.0±254.2</td>
</tr>
<tr>
<td>Nitrites</td>
<td>mg kg⁻¹ Fw</td>
<td>881.5±72.9</td>
<td>1221.6±391.3</td>
<td>2067.9±683.7</td>
<td>3573.9±126.5</td>
<td>1453.0±354.7</td>
</tr>
<tr>
<td>Phosphorus</td>
<td>mg kg⁻¹ Fw</td>
<td>65.0±17.1</td>
<td>437.7±261.4</td>
<td>913.8±353.3</td>
<td>406.5±10.5</td>
<td>96.3±27.9</td>
</tr>
<tr>
<td>Sulfur</td>
<td>mg kg⁻¹ Fw</td>
<td>412.8±59.7</td>
<td>499.7±54.4</td>
<td>579.6±25.7</td>
<td>204.8±14.9</td>
<td>192.8±22.2</td>
</tr>
<tr>
<td>Total phenols</td>
<td>mg GAE kg⁻¹ Fw</td>
<td>3120.8±101.5</td>
<td>3109.9±153.7</td>
<td>1391.5±118.9</td>
<td>1053.8±41.0</td>
<td>2309.7±266.7</td>
</tr>
<tr>
<td>Antioxidant activity</td>
<td>mg TE kg⁻¹ Fw</td>
<td>1033.3±82.0</td>
<td>4015.4±384.7</td>
<td>1311.5±70.5</td>
<td>1592.5±60.4</td>
<td>3333.7±663.0</td>
</tr>
</tbody>
</table>

Fs, fresh weight; GAE, gallic acid equivalents; TE, Trolox equivalents. Values are means±standard errors for each analyzed parameter as determined from n samples (replicates). *Means followed by different letters in each row are significantly different (P<0.05, Tukey’s test).
high chloride and nitrate contents. Finally, cluster 5 comprises S. oleracea, B. officinalis, and D. tenuifolia and is defined by high potassium levels.

To best explain the differences between these different clusters and to evaluate which of the main qualitative parameter(s) play the most important role(s) in determining these differences, linear discriminant analysis was applied. The 13 original variables (i.e., qualitative parameters) of the wild edible plants were reduced to two canonical variates (CN) that represent 81.3% of the total variability in the data (Table 4): 57.7% for the first (CN1) and 23.6% for the second (CN2). To correctly interpret the relationships between CN and the original variables, it is important to note that CN are linear combinations of the original variables and that the canonical coefficients maximise the discrimination among the experimental factors under consideration using canonical coefficients. The original variable with the largest standardised canonical coefficient has the strongest impact on CN. CN1 was positively affected (high standardised canonical coefficient values) by chloride, phosphorus and protein contents and inversely correlated to the levels of dry matter, potassium and fluoride. In contrast, CN2 was mainly defined by phosphorus and calcium levels and the total phenol content (scores of 2.89, 1.49, and 2.47, respectively) and by potassium levels and protein content to a different extent (scores of 1.26 and 0.96, respectively). Coherent information can also be derived from the correlation matrix (Table 4) between the original variables and the new CN. To better interpret the ability of CN1 and CN2 to separate the clusters obtained from the cluster analysis, the selected CN were represented graphically in a biplot (Figure 5), in which the canonical scores were plotted as symbols while the standardised canonical coefficients for each of the original variables were reported as vectors, that was used to interpret the results. According to correlation matrix results (Table 4), the original variables (i.e., sodium, sulfur and antioxidant activity) were not considered in the graphic interpretation because they did not significantly affect the CNj (P<0.05).

Discussion

Wild herbaceous edible plants

This study was found that the quali-quantitative parameters of the wild species examined could make an important contribution to balancing and rationalising diets. In particular, our researcher showed that the edible wild plants contain numerous macro and micronutrients, such as phenols and other components that increase their antioxidant capacity. All the species analysed in the present study, except for B. vulgaris, can be considered low in sodium, which is recommended to reduce blood pressure and prevent heart disease (WHO, 2008). The potassium and magnesium levels are consistent with those reported by Bianco et al. (2009) but are higher than those reported by Renna et al. (2015). While most of calcium values were slightly higher than those reported in the literature (Tarantino et al., 2008). This might be due to the characteristics of the soil in the area, which tend to calcareous, with pH values from 7.7 to 8.2.

The lowest level of nitrates in the present study was observed in the wild C. intybus, which was consistent with the data in the literature (Renna et al., 2015; Bianco, 2002). The highest nitrate accumulation was observed across all the species in the Brassicaceae family (i.e., D. tenuifolia, S. arvensis) and was in agreement with data reported in other studies (Cerruti et al., 1996;...
Santamaria et al., 1998). Moreover, according to the classification proposed by Santamaria (2006), wild C. intybus falls within the group of vegetables considered to have low nitrate contents (200-500 mg kg⁻¹ Fw). Similar values were also seen for P. oleracea, B. vulgaris, S. hispanicus, and F. vulgare.

Data obtained from wild plants (i.e. C. intybus, S. arvensis, and B. officinalis) showed the highest polyphenol contents as well as the greatest antioxidant capacities. In fact, the latter parameter is strictly related to the phenol content as stated by many authors (Di Venere et al., 2009; Sergio et al., 2016).

In general, the qualitative data reported in this study showed that wild edible species contain many of the so-called minor nutrients, such as phenols and antioxidant components, which can further improve an already-balanced diet. This can offer protection against degenerative processes because the wild species considered in this research could make a contribution towards the desired antioxidant capacity of approximately 5000 mg TE kg⁻¹ Fw as estimated by health authorities (INRAN, 2003). Furthermore, in addition to their antioxidant concentrations, all the analysed wild species are rich in phenolic components that increase their antioxidant capacity.
Wild vs cultivated herbaceous food plants

In general, cation and anion contents have previously been reported to vary little among wild and cultivated species. This is probably because the uptake of these components strongly depends on botanical characteristics rather than different environmental and agronomic conditions.

Related to the nitrate contents, the wild species showed significantly lower levels compared to those of cultivated plants (C. intybus, B. officinalis and D. tenuifolia), which have received a certain amount of nitrogen fertilisation. Nitrogen fertilisation is one of the main factors affecting the accumulation of nitrates in vegetables (Santamaria et al., 1999; Weightman et al., 2012).

The total polyphenols and antioxidant activities were significantly higher in the wild plants (C. intybus and B. officinalis) compared to the cultivated plants. Phenol levels vary widely in vegetables depending several factors including the species, physiological stage, and location in terms of the conditions of the soil and the growth environment. In terms of the environment, plants are known to synthesise a greater array of these secondary compounds when they are exposed to several types of stress (e.g., water, saline, thermal, and nutritional) (European Commission, 2002), and wild plants would generally be subjected to greater stress than cultivated plants.

Multivariate analysis of the qualitative parameters

The ANOVA results reported above showed the effects of the experimental factor (i.e., species) on each individual qualitative variable. A multivariate analytical approach (i.e., cluster analysis and linear discriminant analysis) allowed these data to be integrated to evaluate which qualitative variables (considered simultaneously) contributed the most to the differences among the groups (i.e., species).

The graphical results of the linear discriminant analysis show that CN1 is very important for discriminating many of the clusters (especially clusters 2 vs 4). Therefore, the dry matter content; the chloride, phosphorus, potassium, and fluoride levels; and the protein content (i.e., all highly correlated to CN1; Table 4) effectively differentiated between many of these wild edible plant groups. CN2 enables the separation of clusters 2 and 3, showing greater importance than calcium and potassium levels and total phenol content (i.e., all highly correlated to CN2; Table 4). These qualitative parameters are the same as those used to characterised the different clusters in accordance with the ANOVA of the different cluster groups (Table 4).

Conclusions

The present study evaluated the protein, mineral levels polyphenol content and the antioxidant activity of the main wild herbaceous food species consumed in Foglia Province (southern Italy), some of which have not been previously investigated. Most of these analysed wild edible plant species were shown to be good sources of minerals. In particular, S. hispanicus was rich in calcium, potassium and magnesium, D. tenuifolia in phosphorus and sulfur, and C. solstitialis in chlorides.

Nitrates are known to be dangerous at high levels as they can form potentially carcinogenic nitrosamines, but their levels were very low in wild C. intybus compared to the levels in the other species. Among the plants analysed in this study, relatively high, although also variable, total polyphenol contents and antioxidant activity were observed in S. arvensis, C. intybus, B. officinalis, D. erucoides, and S. hispanicus.

For both C. intybus and B. officinalis, significantly higher values of the qualitative parameters were generally observed in wild plants than in cultivated plants. Indeed, wild C. intybus is of particular interest in terms of its qualitative profile, which is characterised by high polyphenol levels and high antioxidant activity. When combined with the low nitrate levels, this profile might account for the high nutritional quality and medicinal properties of

<table>
<thead>
<tr>
<th>Original variable</th>
<th>Standardised canonical coefficient CN1</th>
<th>Standardised canonical coefficient CN2</th>
<th>Pearson's correlation coefficient CN1</th>
<th>Pearson's correlation coefficient CN2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dry matter</td>
<td>-1.99</td>
<td>0.49</td>
<td>-0.66***</td>
<td>-0.54***</td>
</tr>
<tr>
<td>Protein</td>
<td>0.79</td>
<td>-0.86</td>
<td>0.49**</td>
<td>-0.47**</td>
</tr>
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<td>Sodium</td>
<td>0.22</td>
<td>-0.35</td>
<td>0.10ns</td>
<td>-0.10ns</td>
</tr>
<tr>
<td>Potassium</td>
<td>-0.94</td>
<td>-1.26</td>
<td>-0.30*</td>
<td>-0.41**</td>
</tr>
<tr>
<td>Magnesium</td>
<td>1.05</td>
<td>-0.30</td>
<td>0.36*</td>
<td>0.01ns</td>
</tr>
<tr>
<td>Calcium</td>
<td>-0.55</td>
<td>1.49</td>
<td>-0.09ns</td>
<td>0.47**</td>
</tr>
<tr>
<td>Fluorides</td>
<td>-1.69</td>
<td>-0.51</td>
<td>-0.39*</td>
<td>-0.09ns</td>
</tr>
<tr>
<td>Chlorides</td>
<td>4.91</td>
<td>-2.02</td>
<td>0.82***</td>
<td>-0.42**</td>
</tr>
<tr>
<td>Nitrates</td>
<td>0.97</td>
<td>-0.77</td>
<td>0.41**</td>
<td>-0.45**</td>
</tr>
<tr>
<td>Phosphorus</td>
<td>3.23</td>
<td>2.88</td>
<td>0.60***</td>
<td>0.55***</td>
</tr>
<tr>
<td>Sulphur</td>
<td>0.04</td>
<td>-0.23</td>
<td>-0.14ns</td>
<td>-0.14ns</td>
</tr>
<tr>
<td>Total phenols</td>
<td>-0.03</td>
<td>1.47</td>
<td>-0.11ns</td>
<td>0.71***</td>
</tr>
<tr>
<td>Antioxidant activity</td>
<td>0.21</td>
<td>-0.27</td>
<td>0.01ns</td>
<td>-0.27ns</td>
</tr>
<tr>
<td>Percentage of variation explained</td>
<td>57.7%</td>
<td>24.0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cumulative variation explained</td>
<td>-</td>
<td>81.3%</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

CN1, discriminant canonical 1; CN2, discriminant canonical 2. The corresponding percentages of the explained variation are also reported. *P≤0.05; **P≤0.01; ***P≤0.001; ns, correlation not significant (P≥0.05).
the species. Moreover, the value that consumers attribute to wild species for the preparation of particular traditional Italian dishes together with their high organoleptic and nutraceutical levels compared to commonly marketed cultivated species could be the basis for improving the links between rural communities and their plant genetic resources.

References


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