

Changes in soil quality following poplar short-rotation forestry under different cutting cycles

Claudia Di Bene,¹ Elisa Pellegrino,^{1,2} Cristiano Tozzini,¹ Enrico Bonari¹

¹Land Lab, Scuola Superiore Sant'Anna, Pisa; ²Department of Crop Plant Biology, University of Pisa, Italy

Abstract

In the last decade, the change of energy concept induced by global warming and fossil fuel depletion together with the advances in agriculture towards a multifunctional and a more sustainable use of rural areas promoted the development of biomass crops. In this regard, *Populus* is largely utilised in short-rotation forestry (SRF), as it is known to be a fast-growing tree, producing large yields and having a high energy potential. Most studies focused on economic-productive and energetic aspects of *Populus* plantations, whereas their impact on soil quality and health have been poorly investigated. In this study, the main soil chemical parameters, microbial biomass and activity were assessed aiming at evaluating the impact of *Populus* SRF under one, two and three-year cutting cycles (T1, T2 and T3) in comparison with an intensive food cropping system (wheat-soybean rotation, WS). In addition, arbuscular mycorrhizal (AM) fungal inoculum potential was measured using root colonisation (RC) and number of entry points (EP). In the 0-10 cm soil depth, pH, phosphorus (P), total nitrogen (N) and soil organic carbon (SOC) were significantly affected by the management. In comparison with WS, *Populus* SRF treatments produced significant pH decreases together with N and SOC increases, these last ones ranging from 11% to 34% and from 21% to 57%, respectively. Under T3 soil pH decreased of 0.25 units, while P, N and SOC increased of 10%, 34% and 57%, respectively, in comparison with WS. Microbial

biomass and soil respiration under SRF showed also mean increases of 71% and 17%, respectively. Under SRF treatments, *Lolium perenne*, commonly observed in all field plots, was more than twofold colonised by AM fungi in comparison with WS, while the number of EP, observed on *Lactuca sativa* used as a test plant, showed values ranging from 8 to 21 times higher. The present study shows the potential of a *Populus* SRF to improve soil chemical, biochemical and biological quality parameters in comparison with an intensive food cropping system.

Introduction

In the last years, the concept of multifunctional agriculture has obtained large attention from both scientists and policy makers due to its production of both agricultural commodities and ecological services. Such multifunctionality has been focused on an accurate revision of the management of rural areas and of conventional cropping systems both as process and as product (Renting *et al.*, 2009). In this regard, bioenergy crops have recently gained great interest as a potential alternative to agri-food productions and as a clean and renewable energy source reducing greenhouse gas emissions (Wise *et al.*, 2009; Popp *et al.*, 2010). Within biomass crops, *Miscanthus* and *Panicum*, which are perennial rhizomatous grasses, and fast-growing trees, as *Eucalyptus*, *Populus* and *Salix*, are grown worldwide for such bioenergy purposes (Bonari *et al.*, 2004b; Tilman *et al.*, 2006; Karp and Schield, 2008).

Traditionally, *Populus* (poplar) breeding has achieved large success for wood production in short-rotation forestry (SRF) due to the fact that the hybrids of poplar have a fast growth and produce a high yield, although they were not initially selected for growing as coppice (Bonari *et al.*, 2004 a,b; Karp and Schield, 2008). Moreover, many poplar clones and species showed to have a high energy potential and to be able to grow in marginal lands and drought conditions (Hansen, 1991; Makeschin, 1994). Many studies have been performed on biomass productivity and quality, management intensity, economic balance and energy aspects of poplar SRF (Yue *et al.*, 1999; Bonari *et al.*, 2004 a,b; Vande Walle *et al.*, 2007; Karp and Schield, 2008; Lemus *et al.*, 2008; Guidi *et al.*, 2008, 2009; Nassi o Di Nasso, 2010).

So far, less attention has been focused on the evaluation of soil quality changes under SRF management (Tolbert *et al.*, 2002; Kahle *et al.*, 2007; Zornoza *et al.*, 2009; Mao and Zeng, 2010), while many studies were performed on the impact of alternative cropping systems, such as the organic and biodynamic farming, in comparison with high- and low-input conventional managements (Wood and Edwards, 1992; Schjønnning *et al.*, 2002; Hamer *et al.*, 2008; Lagomarsino *et al.*, 2009; Mazzoncini *et al.*, 2010). As regard to soil quality evaluation, Doran and Parkin (1996) proposed a minimum data set of sensitive physical, chemical and biological indicators. Firstly, most studies on soil quality changes under alternative farming utilised chemical and biochemical indicators (Dick, 1983; Kingery *et al.*, 1996; Omay *et al.*, 1997; Carter, 2002), then the biological parameters became more and more impor-

Correspondence: Dr. Claudia di Bene, Land Lab, Scuola Superiore Sant'Anna, p.zza Martiri della Libertà 33, 56127 Pisa, Italy.
Tel: +39.050.883541- Fax: +39.050.883526. E-mail: c.dibene@sssup.it

Key words: arbuscular mycorrhizal fungi, inoculum potential, microbial biomass, root colonisation, short-rotation forestry, soil respiration.

Contributions: CDB and EP contributed equally to this work.

Received for publication: 8 October 2010.
Accepted for publication: 21 December 2010.

©Copyright C. Di Bene *et al.*, 2011
Licensee PAGEPress, Italy
Italian Journal of Agronomy 2011; 6:e6
doi:10.4081/ija.2011.e6

This article is distributed under the terms of the Creative Commons Attribution Noncommercial License (by-nc 3.0) which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

tant in evaluating such changes (Saviozzi *et al.*, 2001; Bending *et al.*, 2004; Parisi *et al.*, 2005; Piotrowski and Rillig, 2008; Mazzoncini *et al.*, 2010). Similarly, evaluating soil quality and health under SRF or following the conversion from food to biomass crops, physical and chemical indicators were largely used (Sartori *et al.*, 2006; Berthrong *et al.*, 2009; Laganère *et al.*, 2010), while recently the attention has been also focused on the biochemical and biological parameters (Makeschin *et al.*, 1994; Guo and Han, 2008; Zornoza *et al.*, 2009; Kahle *et al.*, 2007; 2010; Mao and Zeng, 2010).

The SRF management is associated with minimal mechanical disturbance of the soil and less agrochemical inputs in comparison with conventional cropping systems (Lemus and Lal, 2005; Dickmann, 2006). This, together with the leaf litterfall of deciduous trees, is likely to promote the increase of soil organic carbon (SOC), nitrogen (N) and phosphorus (P) content, as well as of soil microbial biomass (Lal, 2003; Liebig *et al.*, 2005; Ritter, 2007; Iovieno *et al.*, 2010). One of the fundamental components of microbial biomass, which might be affected by SRF, is represented by arbuscular mycorrhizal fungi (AMF) (Rooney *et al.*, 2009). AMF are mutualistic associations between the roots of the majority of plant species and soil borne fungi belonging to Glomeromycota (Schüßler *et al.*, 2001; Smith and Read, 2008). In response to such symbiosis, bioenergy crops could in turn benefit by an increased biomass yield and a greater cropping resistance (Rooney *et al.*, 2009), since AMF are largely known to have a fundamental role in plant nutrition and protection against root and shoot pathogens (Smith and Read, 2008).

The aim of the present study was to evaluate the impact on soil quality of a bioenergy crop management, represented by a SRF poplar plantation under different coppicing frequencies, in comparison with an intensive food cropping system based on a wheat-soybean rotation.

Materials and Methods

Field site and experimental set-up

A long-term poplar (*Populus deltoides* Bartr.) SRF field experiment was started in 1996 at the "Enrico Avanzi" Interdepartmental Centre for Agro-Environmental Research of the University of Pisa (43°40' N lat; 10°19' E long), Italy. Before experimental set-up, the field site was conventionally cultivated with maize (*Zea mays* L.) - durum wheat (*Triticum durum* Desf.) rotation for more than 15 years. The soil showed the following physical and chemical characteristics: clay, 20.1%; silt, 40.5%; sand, 39.4%; available P, 8.8 mg Kg⁻¹; total N, 1.3 g Kg⁻¹; organic carbon, 10.4 g Kg⁻¹. Climatic conditions were typically Mediterranean. More details on climate conditions are given by Mazzoncini *et al.* (2008). The experiment was a completely randomised design, i.e. one, two and three - year cutting cycles (T1, T2 and T3), with three treatments and three replicates (n=3; plots of 500 m²). Details on the poplar stands and their management are given by Nasso *et al.* (2010). In addition, an adjacent intensively tilled (ploughing to 30 cm depth) wheat-soybean rotation (WS), showing similar physical and chemical characteristics in 1996, was selected and used as control. Details on the wheat-soybean experimental design and its management are given by Mazzoncini *et al.* (2008).

Soil and root sampling

In the spring of 2005, one combined soil sample, obtained by mixing three random soil cores, was collected (0-10 cm depth) from each plot and from three random areas within the WS rotation. The soil samples utilised for biochemical analyses were sieved through 2 mm sieve at the field moisture, whereas the samples used for chemical and AMF analyses were oven dried at 30°C before sieving. The root systems of perennial ryegrass (*Lolium perenne* L.), a common weed found in all the plots, known to be

highly responsive to a wide range of AMF, were collected (one combined root sample per each SRF plot and WS area) at a depth of 15 cm and then rinsed and dried (70°C for 3 days).

Analytical procedures

Soil samples were analysed for pH, available P, total N, SOC, microbial biomass carbon (MBC) and soil respiration (SR). Soil pH was measured in deionised water (1:2.5 w/v) (McLean, 1982) and P and N were determined by colorimetry using the Olsen method (Olsen and Sommers, 1982) and by the macro Kjeldahl digestion procedure (Bremner and Mulvaney, 1982), respectively. SOC was evaluated using the modified Walkley-Black wet combustion method (Nelson and Sommers, 1982). MBC was determined by the Vance chloroform fumigation-extraction method, while SR was measured according to the Isermeyer method, described in Alef and Nannipieri (1995). MBC and SR were assessed on soil subsamples of 45 g and SR was determined after 10 days of incubation in closed jars maintained at 25°C. The percentage of AMF colonisation was determined by the gridline-intersect method (Giovannetti and Mosse, 1980) after clearing and staining the roots according to Phillips and Hayman (1970), using lactic acid instead of phenol. AMF infectivity was assessed using the mycorrhizal inoculum potential test (MIP) (Pellegrino *et al.*, 2011) on lettuce (*Lactuca sativa* L.): three seedlings were grown for two weeks in 50 mL sterilised plastic tubes filled with 40 mL of soil obtained by each replicate plot (n=6). Lettuce root system was stained as described above, mounted on microscopic slides and examined under a Reichert-Jung (Vienna, Austria) Polyvar light microscope. The number of entry points (EP) was assessed at a magnification of 125-500x and of 1250x.

Statistical analyses

The soil quality parameters were expressed as percentage of variation in comparison with their values under the intensive wheat management (WS), used as control. For pH values, considering their logarithmic scale, we expressed the variation in units. Data were compared using a one-way (management as factor) analysis of variance (ANOVA). Data were ln- and arcsin-transformed when needed to fulfil the assumptions of ANOVA, which was carried out according to a completely randomised design. The Tukey-B procedure was used to means comparison. Soil chemical parameters showed neither a normal distribution of error terms nor constant error variance, therefore a non-parametric ANOVA was required. In this case, we used the Kruskal-Wallis test and the Mann-Whitney *U*-test as *post-hoc*. All statistics were performed with the SPSS 17.0 software (SPSS Inc., Chicago, IL, USA). Ordination analysis (Redundancy Analysis, RDA) was carried out in Canoco for Windows v. 4.5 (ter Braak and Šmilauer, 2002) in order to investigate the influence of the management (used as explanatory variable) on the soil quality parameters (used as response variables). Additionally, Monte-Carlo permutation test was conducted using 499 random permutations in order to determine the statistical significance.

Results and Discussion

Chemical parameters

In the 0-10 cm soil depth, pH, P, N and SOC were significantly affected by the management (Figure 1). Soil pH, calculated as units of variation in comparison with the value under WS, ranged from -0.26 to -0.13 units in T3 and T1, respectively (Figure 1a). All the poplar SRF treatments produced significant pH decreases in comparison with WS and, within poplar, soil pH significantly decreased from

T1 to T3 (Figure 1a). In a recent meta-analysis, Berthrong *et al.* (2009) reported that *Eucalyptus* and *Pinus* plantations induced a strong and moderate acidification, respectively. A general decrease of pH was also observed in afforestations of a never-tilled soil or grasslands (Ross *et al.*, 1999; Chen *et al.*, 2000; Sartori *et al.*, 2007). Consistently, Guo and Han (2008) reported significant decreases of pH at 0-10 cm and at 10-20 cm soil depth due to soil use conversion based on a 50-year-old *Populus davidiana* plantation. On the contrary, no changes in soil pH were revealed under *Salix* and *Populus* stands in comparison with arable land by Kahle *et al.* (2007, 2010). The reduction of soil pH has been suggested to be related to the higher organic or carbonic acid production, the latter due to an increased autotrophic respiration (Richter and Markewitz, 1995) and to the influence of tree root system on level of ground water and cation uptakes (Attiwell and Adams, 1993; Jobbágy and Jackson, 2003). Here, the differences in acidification among the different SRF treatments may be due to the larger production of tree root biomass, under less frequent cutting cycles, that may release a higher number of H⁺ ions (Attiwell and Adams, 1993).

The variation of soil available P ranged from -17% to 10% in T1 and T3, respectively (Figure 1b). Significant soil P changes were observed between T1-T2 in comparison with WS and within poplar SRF, T3 showed a significant soil P increase, in comparison with T1 and T2. Some authors reported increases of soil P due to the afforestation with different tree species in comparison with grasslands or agricultural soils (Ritter, 2007; Zornoza *et al.*, 2009), while some others observed higher P contents or no changes in arable lands or pastures than in adjacent forests (Koerner *et al.*, 1997; Ross *et al.*, 1999; Chen *et al.* 2000; Zhao *et al.*, 2007). Such contrasting results might be due to variables influencing P dynamics, which may be associated with SOC changes as reported by Piccolo *et al.* (1996), and to previous land-use, time since land-use conversion, tree species planted and climatic conditions (Ross *et al.*, 1999; Ritter, 2007; Zhao *et al.*, 2007).

The total soil N variations ranged from 11% to 34% in T2 and T3, respectively (Figure 1c). All poplar SRF showed significant soil N increases in comparison with WS and, within such treatments, T3 produced a significantly higher increase than T1-T2 (Figure 1c). Consistently with our data, in other studies soil N concentration in the shallow layer was lower in agricultural lands than in *Betula* and *Larix* thinned closed canopy plantations and in *Populus* stands (Ritter *et al.*, 2007; Sartori *et al.*, 2007). Lower soil N was also detected in agricultural lands than in deciduous forests by Morris *et al.* (2007). By contrast, soil N decreases were reported under *Pinus* and *Eucalyptus* afforestations (Binkley and Resh, 1999; Berthrong *et al.*, 2009).

The change of SOC due to the different treatments in comparison with WS ranged from 21% to 57% in T1 and T3, respectively (Figure 1d). A significant increase was observed under all poplar SRF treatments and within poplar stands, SOC under T3 was significantly higher than under T1 and T2 (Figure 1d). The increase of SOC under SRF may result in positive changes of soil structure, water retention, nutrient availability, biological diversity and C sequestration, since it is well known to affect directly or indirectly the overall soil quality parameters (Schjønning *et al.*, 2004). Here, SOC showed a pattern similar to the soil N concentration as reported in other studies (Franzluebbers and Stuedemann, 2009; Yao *et al.*, 2010). Such similarity between SOC and N patterns was previously explained by carbon inputs from plant production and outputs through microbial decomposition (Gill *et al.*, 1999). Along with our results, several studies observed SOC increases due to the afforestation of agricultural soils (Park *et al.*, 1994; Grigal and Berguson, 1998; Tolbert *et al.*, 2002; Kahle *et al.*, 2007; Laganière *et al.*, 2010), while some others reported no changes or lower SOC concentration under forest than under adjacent grassland (Berthrong *et al.*, 2009; Chen *et al.*, 2010;

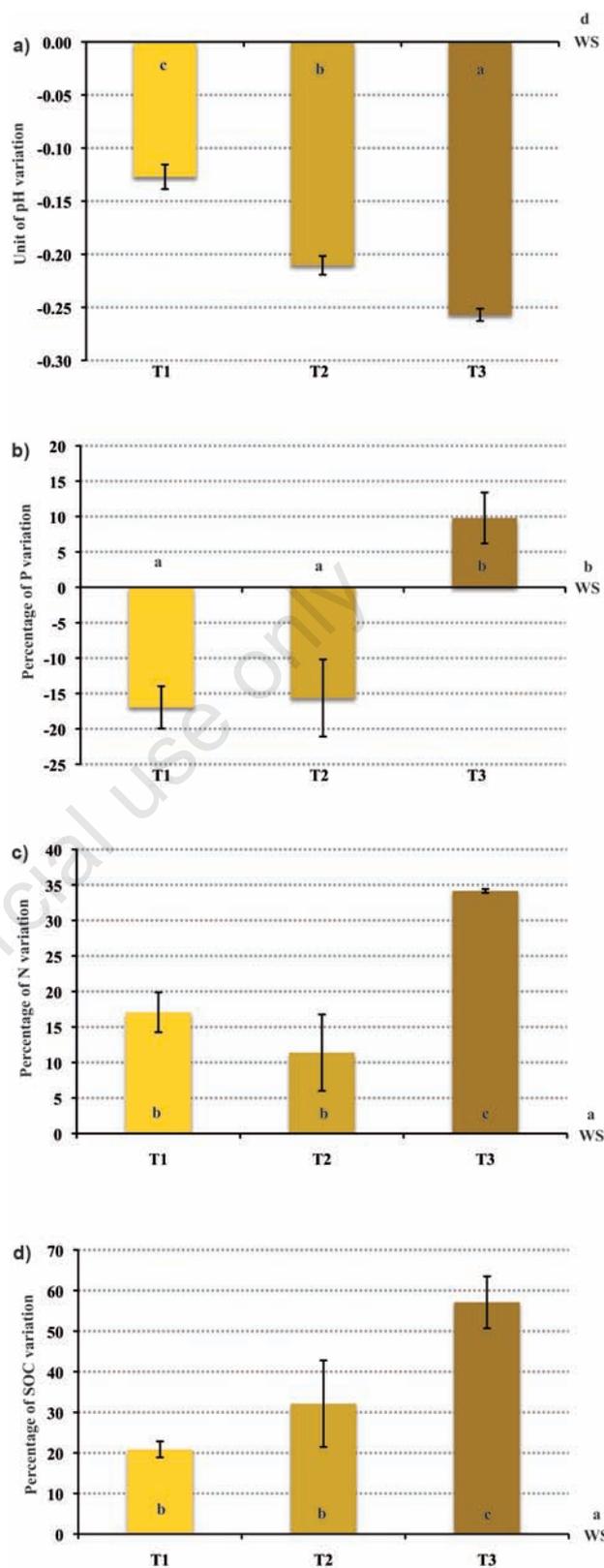


Figure 1. Soil pH a), available phosphorus (P); b), total nitrogen (N); c) and organic carbon (SOC); d) under poplar short-rotation forestry (one, two and three-year cutting cycles: T1, T2 and T3). The values are expressed as unit or percentage of variation in comparison with their values under a wheat-soybean rotation (WS). Different letters indicate significant differences as tested by the Kruskal-Wallis test ($P \leq 0.05$) and the Mann-Whitney *U*-test as *post-hoc* ($P \leq 0.05$).

Mao and Zeng, 2010). Climate, soil type, management, land use and time since land use conversion may explain these contrasting results (Paul *et al.*, 2002; Laganière *et al.*, 2010).

Biochemical parameters

Most studies have used biochemical indicators, such as the MBC and the SR, aiming to evaluate the impact of different managements on soil quality (Haynes, 1999; Dilly and Nannipieri, 2001; Lagomarsino *et al.*, 2009; Iovieno *et al.*, 2010), while under SRF such parameters have been less investigated (Guo and Han, 2008; Mao and Zeng, 2010). Here, MBC and SR showed to be significantly affected by the management (Figure 2). MBC was significantly increased by all poplar SRF treatments compared with WS (Figure 2a), showing variations from 43% to 93% (T1 and T3, respectively). Our results are in agreement with other studies evaluating the effects of agricultural land conversion to *Populus*, *Quercus* and *Salix* plantations on MBC (Makeschin, 1994; Zornoza *et al.*, 2009; Kahle *et al.*, 2010; Mao and Zeng, 2010). By contrast, other authors showed no changes or significant decreases of MBC under *Pinus* stands (Chen *et al.*, 2000; Macdonald *et al.*, 2009). The MBC increases observed here and in other studies may be explained by the increase in carbon available for microorganisms derived from rhizodeposition and from the high-quality litter of *Salicaceae* and *Fagaceae*, while the lower soil MBC under *Pinus* afforestation, as compared to the soil under the climax vegetation, was attributed to the low-quality litter of pine needle litter by Iovieno *et al.* (2010). In addition, the mean decrease of number of live bacteria in the soils amended with *Pinus* in comparison with *Quercus* observed by Grenni *et al.* (2009) may contribute to explain

the differences of MBC changes commonly reported between *Pinus* and other trees.

SR percentages of variation ranged from 8% to 25% in T1 and T3, respectively (Figure 2b). SR values under T2 and T3 were significantly higher than that under WS and, within the different cutting cycles, T2-T3 and T1 produced significantly different effects on SR (T2-T3>T1) (Figure 2b). According to our data, Zornoza *et al.* (2009), studying the impact of different land use, observed higher values of SR under forest than under abandoned and agricultural systems. The SR pattern, similar to the MBC one, may be explained by the higher quantity and different quality of litter under the tree stands in comparisons with herbaceous-based systems (Singh and Singh, 1995; Chen *et al.*, 2000).

Arbuscular mycorrhizal fungi measurements

L. perenne, the common plant species found in all the plots, showed root colonisation (RC) changes ranging from 141% to 170% in T1 and T2, respectively (Figure 3a). *L. perenne* grown under WS was significantly less colonised by AMF than that grown under poplar SRF and, within the different cutting cycles, the root colonisation under T1 and T3 was significantly lower than that under T2 (Figure 3a). The difference of AMF colonisation between SRF and WS may be attributed to the cultural operations carried out in order to prepare seedbed, to fertilise crops and to control weeds, pests and diseases as well as to the above- and belowground plant species diversity (Helgason *et al.*, 1998; Vandenkoornhuyse *et al.*, 2003; Leake *et al.*, 2004). In addition, the highest root colonisation of the *L. perenne* grown under T2 might be explained by a large production of poplar fine roots observed in such management (Amato, 2000; Bonari and Masoni, 2000).

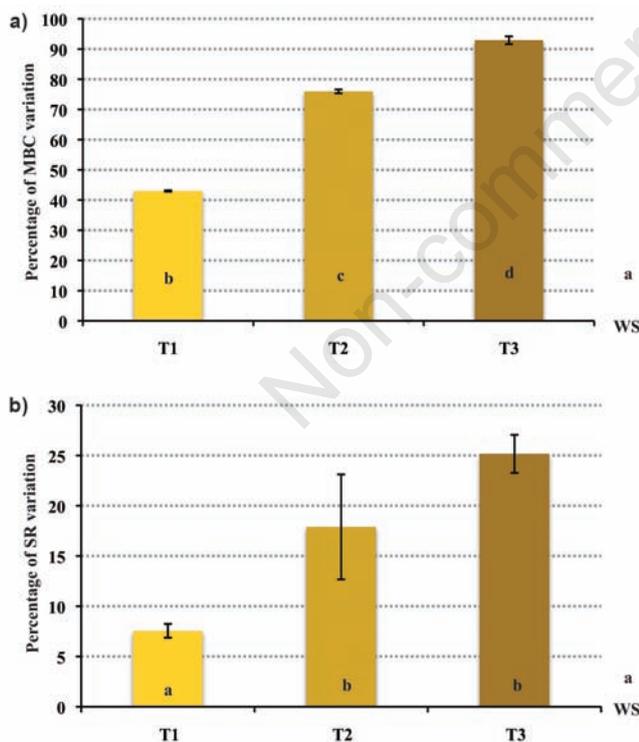


Figure 2. Soil microbial biomass (MBC) a) and soil respiration (SR) b) under poplar short-rotation forestry (one, two and three-year cutting cycles: T1, T2 and T3). The values are expressed as percentage of variation in comparison with their values under a wheat-soybean rotation (WS). Different letters indicate significant differences as tested by ANOVA ($P \leq 0.001$) and the Tukey-B test as *post-hoc*.

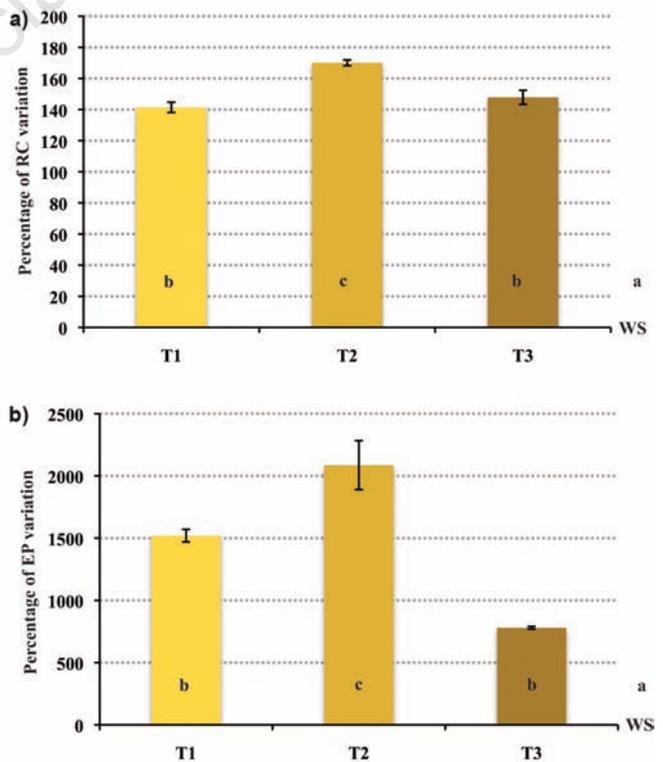


Figure 3. Arbuscular mycorrhizal fungal root colonisation (RC), a) and number of entry points (EP) b) under poplar short-rotation forestry (one, two and three-year cutting cycles: T1, T2 and T3). The values are expressed as percentage of variation in comparison with their values under a wheat-soybean rotation (WS). Different letters indicate significant differences as tested by ANOVA ($P \leq 0.001$) and the Tukey B test as *post-hoc*.

The number of EP under poplar SRF treatments showed values from 8 to 21 times higher than that reported under WS (Figure 3b). The number of EP under T1 and T3 was significantly lower than under T2 ($T1-T3 < T2$) (Figure 3b). Our EP data under WS are consistent with the values reported by several authors, which measured such parameter assessing the AMF inoculum potential under shrubs, wild and cultivated plant species from semiarid ecosystem to boreal grasslands (Requena *et al.*, 2001; Bharadwaj *et al.*, 2007). Besides, EP values similar to those reported, here, under SRF were observed for different AMF inocula on several plant species (Liu and Luo, 1994). Such strong difference of AMF inoculum potential between a herbaceous-based system (WS) and the SRF may be explained by the different management, plant communities, patterns of root systems and hyphal networks, AMF communities in the soil and *in planta* (Bever *et al.*, 1996; Helgason *et al.*, 1998; van der Heijden *et al.*, 1998; Daniell *et al.*, 2001; Vandenkoornhuyse *et al.*, 2003; Giovannetti *et al.*, 2004).

Multivariate analysis of the soil chemical, biochemical and biological parameters

The RDA analysis, aiming at evaluating the impact of the different managements on the soil quality parameters, showed that management, used as explanatory variable, explained 69.9% (I and II axes) of the whole variance and that its effect on soil quality parameters (Figure 4), used as response variables, was significant ($P=0.002$). In detail, the Monte-Carlo permutation test pointed out significant differences on soil quality between WS and poplar SRF stands ($P=0.01$) and between T3 and the other cutting cycles ($P=0.002$), as showed by the distances of the centroids representing the managements. The biplot shows that the values of all soil quality parameters were higher under poplar SRF in comparison with WS, and that the differences among the T1, T2 and T3 were due to the fact that T1 and T2 increased the parameters linked to AMF, while T3 the soil chemical and biochemical variables. The short distance between the arrows representing RC and EP, as well as those representing SOC and N, shows the strong correlation between such parameters.

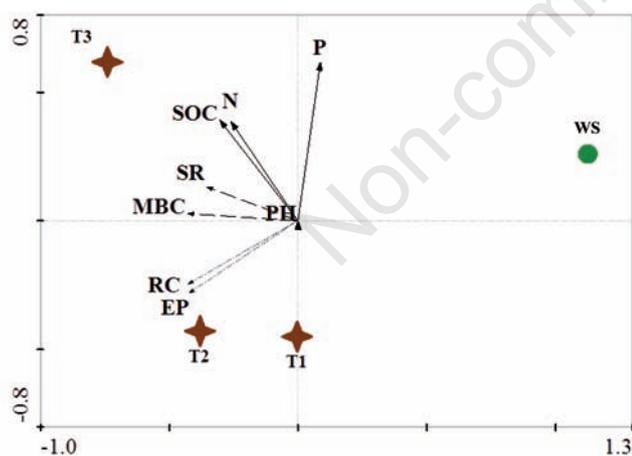


Figure 4. Redundancy Analysis (RDA) biplot based on: chemical, biochemical and AMF parameters (pH; P, available phosphorus; N, Kjeldahl nitrogen; SOC, soil organic carbon; SR, soil respiration; MBC, microbial biomass carbon; RC, mycorrhizal colonization of *Lolium perenne*; EP, number of entry points) and treatments (one, two and three-year cutting cycles poplar short-rotation forestry: T1, T2 and T3, respectively; wheat-soybean rotation, WS). Treatments are represented by stars (T1, T2 and T3) and circle (WS). The chemical, biochemical and AMF parameters are represented by arrows. The 1st and 2nd axis accounted for 49.0% and 69.9% of the variability explained by all canonical axes and were significant ($P=0.002$).

Conclusions

Since biomass is one of the most important sources of renewable energy, plant-microbial interactions under *Poplar* stands in comparison with conventional agricultural management are a cutting-edge issue. The present study shows the potential of a bioenergy crop management, represented by poplar SRF, to improve soil quality in comparison with an intensive food cropping system and a distinct behaviour of the different poplar cutting cycles in promoting soil organic carbon, microbial biomass and AMF inoculum potential. Such findings have important ecological and environmental implications, since the positive belowground effects observed here under poplar plantations could improve the viability of low-input SRF stands. The interactions between bioenergy crops and microorganisms need to be further investigated to explore their implications on plant-soil carbon sequestration, biomass production and nutrient uptake by mycorrhizas.

References

- Alef K., Nannipieri P., 1995. Methods in Applied Soil Microbiology and Biochemistry. Academic Press, London, UK.
- Amato M., 2000. Parametri che definiscono l'accrescimento radicale. In: G. Mosca and T. Vamerali (eds.) Obiettivo radice metodi di studio e risultati ottenuti in ambiente mediterraneo. Cleup Publ., Padova, Italy, pp 73-82.
- Attiwell P.M., Adams M.A., 1993. Nutrient cycling in forests. New Phytol., 124:561-582.
- Bending G.D., Turner M.K., Rayns F., Marx M.C., Wood M., 2004. Microbial and biochemical soil quality indicators and their potential for differentiating areas under contrasting agricultural management regimes. Soil Biol. Biochem. 36:1785-1792.
- Berthrong S.T., Jobbágy E.G., Jackson R.B., 2009. A global meta-analysis of soil exchangeable cations, pH, carbon, and nitrogen with afforestation. Ecol. Appl. 19:2228-2241.
- Bever J.D., Morton J.B., Antonovics J., Schultz P.A., 1996. Host-dependent sporulation and species diversity of arbuscular mycorrhizal fungi in a mown grassland. J. Ecol. 84:71-82.
- Bharadwaj D.P., Lundquist P.O., Alstrom S., 2007. Impact of plant species grown as monocultures on sporulation and root colonization by native arbuscular mycorrhizal fungi in potato. Appl. Soil Ecol. 35:213-225.
- Binkley D., Resh S., 1999. Rapid changes in soils following Eucalyptus afforestation in Hawaii. Soil. Sci. Soc. Am. J. 63:222-225.
- Bonari E., Masoni A., 2000. Radice sinonimo di sostanza organica per la rizosfera. In: G. Mosca and T. Vamerali (eds.) Obiettivo radice metodi di studio e risultati ottenuti in ambiente mediterraneo. Cleup Publ., Padova, Italy, pp 44-46.
- Bonari E., Picchi G., Ginanni M., Fraga A., Guidi W., 2004a. Poplar short rotation coppice behaviour under different harvesting treatments. In: W.P.M. Van Swaaij, T. Fjällström, P. Helm and A. Grassi (eds.) Second World Biomass Conference. ETA Publ., Firenze, Italy, pp 237-239.
- Bonari E., Picchi G., Ginanni M., Guidi W., Piccioni E., Fraga A., Villani R. 2004b. Le colture da energia. In: A. Faini and G. Nocentini (eds.) Le colture dedicate ad uso energetico: il progetto Bioenergy Farm. ARSIA Publ., Firenze, Italy, pp 29-78.
- Bremner J.M., Mulvaney C.S., 1982. Nitrogen - Total. In: A.L. Page, R.H. Miller and D.R. Keeney (eds.) Methods of Soil Analysis, Part 2, Chemical and microbiological properties, second edition. Agronomy Monograph 9, American Society of Agronomy, Madison, WI, USA, pp 595-624.

- Carter M.R., 2002. Soil quality for sustainable land management: organic matter and aggregation interactions that maintain soil functions. *Agron. J.* 94:38-47.
- Chen C.R., Condon L.M., Davis M.R., Sherlock R.R., 2000. Effects of afforestation on phosphorus dynamics and biological properties in a New Zealand grassland soil. *Plant Soil* 220:151-163.
- Chen Y., Olson D.M., Ruberson J.R., 2010. Effects of nitrogen fertilization on tritrophic interactions. *Arthropod Plant Interact.* 4:81-94.
- Daniell T.J., Husband R., Fitter A.H., Young J.P.W., 2001. Molecular diversity of arbuscular mycorrhizal fungi colonising arable crops. *Fems Microbiol. Ecol.* 36:203-209.
- Dick W.A., 1983. Organic carbon, nitrogen and phosphorus concentrations and pH in soil profiles as affected by tillage intensity. *Soil Sci. Soc. Am. J.* 47:102-107.
- Dickmann D.L., 2006. Silviculture and biology of short-rotation woody crops in temperate regions: Then and now. *Biomass Bioenerg.* 30:696-705.
- Dilly O., Nannipieri P., 2001. Response of ATP content, respiration rate and enzyme activities in an arable and a forest soil to nutrient additions. *Biol. Fert. Soils* 34:64-72.
- Doran J.W., Parkin T.B., 1996. Quantitative indicators of soil quality: a minimum data set. In: J.W. Doran and A.J. Jones (eds) *Methods for Assessing Soil Quality*. SSSA, Inc. Publ., Madison, WI, USA, pp 25-37.
- Franzluebbers A.J., Stuedemann J.A., 2009. Soil-profile organic carbon and total nitrogen during 12 years of pasture management in the Southern Piedmont USA. *Agr. Ecosyst. Environ.* 129:28-36.
- Gill R., Burke I.C., Milchunas D.G., Lauenroth W.K., 1999. Relationship between root biomass and soil organic matter pools in the shortgrass steppe of Eastern Colorado. *Ecosystems* 2:226-236.
- Giovannetti M., Mosse B., 1980. An evaluation of techniques for measuring vesicular-arbuscular mycorrhizal infection in roots. *New Phytol.* 84:489-500.
- Giovannetti M., Sbrana C., Avio L., Strani P., 2004. Patterns of below-ground plant interconnections established by means of arbuscular mycorrhizal networks. *New Phytol.* 164:175-181.
- Grenni P., Barra Caracciolo A., Rodriguez-Cruz M.S., Sanchez-Martin M.J., 2009. Changes in the microbial activity in a soil amended with oak and pine residues and treated with linuron herbicide. *Appl. Soil Ecol.* 41:2-7.
- Grigal D.F., Berguson W.E., 1998. Soil carbon changes associated with short-rotation systems. *Biomass Bioenerg.* 14:371-377.
- Guidi W., Piccioni E., Ginanni M., Bonari E., 2008. Bark content estimation in poplar (*Populus deltoides*) short-rotation coppice in central Italy. *Biomass Bioenerg.* 33:1703-1709.
- Guidi W., Tozzini C., Bonari E., 2009. Estimation of chemical traits in poplar short-rotation coppice at stand level. *Biomass Bioenerg.* 32:518-524.
- Guo Y.J., Han J.G., 2008. Soil biochemical properties and arbuscular mycorrhizal fungi as affected by afforestation of rangelands in northern China. *J. Arid Environ.* 72:1690-1967.
- Hamer U., Makeschin F., Stadler J., Klotz S., 2008. Soil organic matter and microbial community structure in set-aside and intensively managed arable soils in NE-Saxony, Germany. *Appl. Soil Ecol.* 40:465-475.
- Hansen E.A., 1991. Poplar woody biomass yields: a look to the future. *Biomass Bioenerg.* 1:1-7.
- Haynes R.J., 1999. Size and activity of the soil microbial biomass under grass and arable management. *Biol. Fert. Soils* 30:210-216.
- Helgason T., Daniell T.J., Husband R., Fitter A.H., Young J.P.W., 1998. Ploughing up the wood-wide web? *Nature* 394:431.
- Iovieno P., Alfani A., Bååth E., 2010. Soil microbial community structure and biomass as affected by *Pinus pinea* plantation in two Mediterranean areas. *Appl. Soil Ecol.* 45:56-63.
- Jobbágy E.G., Jackson R.B., 2003. Patterns and mechanisms of soil acidification in the conversion of grasslands to forests. *Biogeochemistry* 64:205-229.
- Kahle P., Baum C., Boelcke B., Kohl J., Ulrich R., 2010. Vertical distribution of soil properties under short-rotation forestry in Northern Germany. *J. Plant Nutr. Soil Sci.* 173:737-746.
- Kahle P., Hildebrand E., Baum C., Boelcke B., 2007. Long-term effects of short rotation forestry with willows and poplar on soil properties. *Arch. Acker. Pfl. Boden.* 53:673-682.
- Karp A., Shield I., 2008. Bioenergy from plants and the sustainable yield challenge. *New Phytol.* 179:16-32.
- Kingery W.L., Wood C.W., Williams J.C. 1996. Tillage and amendment effects on soil carbon and nitrogen mineralization and phosphorus release. *Soil Tillage Res.* 37:239-250.
- Koerner W., Dupouey J.L., Dambrine E., Benoît M., 1997. Influence of past land use on the vegetation and soils of present day forest in the Vosges mountains, France. *J. Ecol.* 85:351-358.
- Laganière J., Angers D.A., Paré D., 2010. Carbon accumulation in agricultural soils after afforestation: a meta-analysis. *Global Change Biol.* 16:439-453.
- Lagomarsino A., Moscatelli M.C., Di Tizio A., Mancinelli R., Grego S., Marinari S., 2009. Soil biochemical indicators as a tool to assess the short-term impact of agricultural management on changes in organic C in a Mediterranean environment. *Ecologic. Indicat.* 9:518-527.
- Lal R., 2003. Global potential of soil carbon sequestration to mitigate the greenhouse effect. *Crit. Rev. Plant Sci.* 22:151-184.
- Leake J.R., Johnson D., Donnelly D.P., Muckle G.E., Boddy L., Read D.J., 2004. Networks of power and influence: the role of mycorrhizal mycelium in controlling plant communities and agroecosystem functioning. *Can. J. Botany* 82:1016-1045.
- Lemus R., Brummer E.C., Burras C.L., Moore K.J., Barker M.F., Molstad N.E., 2008. Effects of nitrogen fertilization on biomass yield and quality in large fields of established switchgrass in southern Iowa, USA. *Biomass Bioenerg.* 32:1187-1194.
- Lemus R., Lal R., 2005. Bioenergy crops and carbon sequestration. *Cr. Rev. Plant Sci.* 24:1-21.
- Liebig M.A., Morgan J.A., Reeder J.D., Ellert B.H., Gollany H.T., Schuman G.E., 2005. Greenhouse gas contributions and mitigation potential of agriculture practices in northwestern USA and western Canada. *Soil Till. Res.* 83:25-52.
- Liu R. J., Luo X. S. 1994. A new method to quantify the inoculum potential of arbuscular mycorrhizal fungi. *New Phytol.*, 128:89-92.
- Macdonald C.A., Thomas N., Robinson L., Tate K.R., Ross D.J., Dando J., Singh B.K., 2009. Physiological, biochemical and molecular response of the soil microbial community after afforestation of pastures with *Pinus radiata*. *Soil Biol. Biochem.* 41:1642-1651.
- Makeschin F., 1994. Effects of energy forestry on soils. *Biomass Bioenerg.* 6:63-79.
- Mao R., Zeng D.H., 2010. Changes in soil particulate organic matter, microbial biomass, and activity following afforestation of marginal agricultural lands in a semi-arid area of Northeast China. *Environ. Manage.* 46:110-116.
- Mazzoncini M., Canali S., Giovannetti M., Castagnoli M., Tittarelli F., Antichi D., Nannelli R., Cristani C., Bàrberi P., 2010. Comparison of organic and conventional stockless arable systems: A multidisciplinary approach to soil quality evaluation. *Appl. Soil Ecol.* 44:124-132.
- Mazzoncini M., Di Bene C., Coli A., Antichi D., Petri M., Bonari E., 2008. Rainfed wheat and soybean productivity in a long-term tillage experiment in central Italy. *Agron. J.* 100:1418-1429.
- McLean E.O., 1982. Soil pH and lime requirement. In: A.L. Page, R.H. Miller and D.R. Keeney (eds.) *Methods of Soil Analysis, Part 2*,

- Chemical and microbiological properties, second edition. Agronomy Monograph 9, American Society of Agronomy, Madison, WI, USA, pp 199-224.
- Morris S.J., Bohm S., Haile-Mariam S., Paul E.A., 2007. Evaluation of carbon accrual in afforested agricultural soils. *Glob. Change Biol.* 13:1145-1156.
- Nassi o Di Nasso N., Guidi W., Ragaglini G., Tozzini C., Bonari E., 2010. Biomass production and energy balance of a 12-year old short-rotation coppice poplar stand under different cutting cycles. *Glob. Change Biol. Bioenerg.* 2:89-97.
- Nelson D.W., Sommers L.E., 1982. Total carbon, organic carbon and organic matter. A.L. Page, R.H Miller and D.R. Keeney (eds.) *Methods of Soil Analysis, Part 2, Chemical and microbiological properties, second edition.* Agronomy Monograph 9, American Society of Agronomy, Madison, WI, USA, pp 539-579.
- Olsen S.R., Sommers L.E., 1982. Phosphorus. In: A.L. Page, R.H Miller and D.R. Keeney (eds.) *Methods of Soil Analysis, Part 2, Chemical and microbiological properties, second edition.* Agronomy Monograph 9, American Society of Agronomy, Madison, WI, USA, pp 403-430.
- Omay A.B., Rice C.W., Maddux L.D., Gordon W.B., 1997. Changes in soil microbial and chemical properties Under long-term crop rotation and fertilization. *Soil Sci. Soc. Am. J.* 61:1672-1678.
- Parisi V., Menta C., Gardi C., Jacomini C., Mozzanica E., 2005. Microarthropod communities as a tool to assess soil quality and biodiversity: A new approach in Italy. *Agr. Ecosyst. Environ.* 105:323-333.
- Park J., Newman S.M., Cousins S.H. 1994. The effects of poplar (*Populus deltoides*) on soil properties in a silvoarable system. *Agroforest. System.* 25:111-118.
- Paul K.I., Polglase P.J., Nyakuengama J.G., Khanna P.K., 2002. Change in soil carbon following afforestation. *For. Ecol. Manage.* 168:241-257.
- Pellegrino E., Bedini S., Avio L., Bonari E., Giovannetti, M., 2011. Field inoculation effectiveness of native and exotic arbuscular mycorrhizal fungi in a Mediterranean agricultural soil. *Soil Biol. Biochem.* 43:367-376.
- Phillips J.M., Hayman D.S., 1970. Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. *T. Brit. Mycol. Soc.* 55:158-161.
- Piccolo A., Nardi S., Concheri G., 1996. Macromolecular changes of humic substances induced by interaction with organic acids. *Eur. J. Soil Sci.* 47:319-328.
- Piotrowski J.S., Rillig M.C., 2008. Succession of arbuscular mycorrhizal fungi: patterns, causes, and considerations for organic agriculture. *Adv. Agron.* 97:111-130.
- Popp A., Lotze-Campen H., Leimbach M., Knof B., Beringer T., Bauer N., Bodirsky B., 2010. On sustainability of bioenergy production: integrating co-emissions from agricultural intensification. *Biomass Bioenerg.* doi:10.1016/j.biombioe.2010.06.014.
- Renting H., Rossing W.A.H., Groot J.C.J., Van der Ploeg, J.D., Laurent C., Perraud D., Stobbelaar D.J., Van Ittersum M.K., 2009. Exploring multifunctional agriculture. A review of conceptual approaches and prospects for an integrative transitional framework. *J. Environ. Manage.* 90:112-123.
- Requena N., Pérez-Solis E., Azcón-Aguilar C., Jeffries P., Barea J.M., 2001. Management of indigenous plant-microbe symbioses aids restoration of desertified ecosystems. *Appl. Environ. Microb.* 67:495-498.
- Richter D.D., Markewitz D., 1995. How deep is soil? *BioScience*, 45:600-609.
- Ritter E., 2007. Carbon, nitrogen and phosphorus in volcanic soils following afforestation with native birch (*Betula pubescens*) and introduced larch (*Larix sibirica*) in Iceland. *Plant Soil* 295:239-251.
- Rooney, D.C., Killham, K., Bending, G.D., Baggs, E., Weih, M., Hodge, A., 2009. Mycorrhizas and biomass crops: opportunities for future sustainable development. *Trends Plant Sci.* 14:542-549.
- Ross D., Tate K., Scott N., Feltham C., 1999. Land-use change: effects on soil carbon, nitrogen and phosphorus pools and fluxes in three adjacent ecosystems, *Soil Biol. Biochem.* 31:803-813.
- Sartori F., Lal R., Ebinger M.H., Eaton J.A., 2007. Changes in soil carbon and nutrient pools along a chronosequence of poplar plantations in the Columbia Plateau, Oregon, USA. *Agr. Ecosyst. Environ.* 122:325-339.
- Sartori F., Lal R., Ebinger M.H., Parrish D.J., 2006. Potential soil carbon sequestration and CO₂ offset by dedicated energy crops in the USA. *Crit. Rev. Plant Sci.* 25:441-472.
- Saviozzi A., Levi-Minzi R., Cardelli R., Riffaldi R., 2001. A comparison of soil quality in adjacent cultivated, forest and native grassland soils. *Plant Soil* 233:251-259.
- Schjøning, P., Elmholt, S., Christensen, B.T., 2004. Managing soil quality: challenges in modern agriculture. CABI Publ., Wallingford, UK.
- Schjøning P., Munkholm L.J., Moldrup P., Jacobsen O.H., 2002. Modelling soil pore characteristics from measurements of air exchange: the long-term effects of fertilization and crop rotation. *Eur. J. Soil Sci.* 53:331-339.
- Schüßler A., Schwarzott D., Walker C., 2001. A new fungal phylum, the Glomeromycota: phylogeny and evolution. *Mycol. Res.* 105:1413-1421.
- Singh, S., Singh J.S., 1995. Microbial biomass associated with water-stable aggregates in forest, savanna and cropland soils of a seasonally dry tropical region, India. *Soil Biol. Biochem.* 27:1027-1033.
- Smith S.E., Read D.J., 2008. *Mycorrhizal symbiosis.* 3rd edition. Academic Press, Amsterdam, The Netherlands.
- ter Braak C.J.F., Šmilauer P., 2002. *CANOCO Reference manual and CanoDraw for Windows User's guide: Software for canonical community ordination (version 4.5).* Microcomputer Power, Ithaca, NY, USA.
- Tilman D., Hill J., Lehman C., 2006. Carbon-negative biofuels from low-input high-diversity grassland biomass. *Science* 314:1598-1600.
- Tolbert V.R., Todd Jr D.E., Mann L.K., Jawdy C.M., Mays D.A., Malik R., Bandaranayake W., Houston A., Tyler D., Pettry D.E., 2002. Changes in soil quality and below-ground carbon storage with conversion of traditional agricultural crop lands to bioenergy crop production. *Environ. Pollut.* 116:S97-S106.
- van der Heijden M.G.A., Klironomos J.N., Ursic M., Moutoglis P., Streitwolf-Engel R., Boller T., Wiemken A., Sanders I.R., 1998. Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. *Nature* 396:69-72.
- Vande Walle I., Van Camp N., Van de Castele L., Verheyen K., Lemeur R., 2007. Short-rotation forestry of birch, maple, poplar and willow in Flanders (Belgium) I. Biomass production after 4 years of tree growth. *Biomass Bioenerg.*, 31:267-275.
- Vandenkoornhuyse P., Ridgway K.P., Watson I.J., Fitter A.H., Young J.P.W., 2003. Co-existing grass species have distinctive arbuscular mycorrhizal communities. *Mol. Ecol.* 12:3085-3095.
- Wise M., Calvin K., Thomson A., Clarke L., Bond-Lamberty B., Sands R., Smith S.J., Janetos A., Edmonds J., 2009. Implications of limiting CO₂ concentrations for land use and energy. *Science* 324:1183-1186.
- Wood C.W., Edwards J.H., 1992. Agroecosystem management effects on soil carbon and nitrogen. *Agr. Ecosyst. Environ.* 39:123-138.
- Yao Z., Zhou Z., Zheng X., Xie B., Mei B., Wang R., Butterbach-Bahl K.,

- Zhu J., 2010. Effects of organic matter incorporation on nitrous oxide emissions from rice-wheat rotation ecosystems in China. *Plant Soil* 327:315-330.
- Yue M., Ren Y., Dang G.D., Gu T.Q., 1999. Species diversity of higher plant communities in Foping Natural Reserve. *Chinese Biodivers.* 7:263-269.
- Zhao Q., Zeng D.H., Lee D.K., He X.Y., Fan Z.P., Jin Y.H., 2007. Effects of *Pinus sylvestris* var. *mongolica* afforestation on soil phosphorus status of the Keerqin Sandy Lands in China. *J. Arid Environ.* 69:569-582.
- Zornoza R., Mataix-Solera J., Guerrero C., Arcenegui V., Mataix-Beneyto J., 2009. Comparison of soil physical, chemical, and biochemical properties among native forest, maintained and abandoned almond orchards in mountainous areas of Eastern Spain. *Arid Land Res. Manag.* 23:267-282.

Non-commercial use only